# Nutrient Concentration Effects on *Pisolithus tinctorius* Development on Containerized Loblolly Pine (*Pinus taeda* L.) Seedlings

J. L. Torbert, J. A. Burger, and R. E. Kreh

Research assistant, associate professor, and research associate, Department of Forestry, Virginia Polytechnic Institute and State University, Blacksburg

A trade-off between seedling size and degree of mycorrhizal infection needs to be considered when determining the rate at which to fertilize containerized seedlings infected with ectomycorrhizae. Tree Planters' Notes 37(3):17-22; 1986.

Ectomycorrhizae can increase forest tree survival and growth under many different site conditions (11). Mycorrhizae increase tree root surface area, increase the absorption of water and nutrients, and can exploit forms of phosphorus (P) unavailable to nonmycorrhizal plants (1). Pisolithus tinctorius (Pers.) Coker & Couch is a fungus capable of forming ectomycorrhizae on trees growing on acidic, droughty, and infertile soils. It has been used successfully for improving the performance of pines on reclaimed surface mines (6) and on clay soils in North Carolina and Georgia (4).

Much work has been done on methods for inoculating pines with *P. tinctorius* basidiospores (5, 8, 9). Using spores is more convenient and less costly than inoculating with vegetative fungal mycelia. Marx and co-workers (7) found that ectomycorrhizal development was

related to soil fertility, with nitrogen (N) and P being the most critical nutrient elements. At high N and P concentrations, P. tinctorius infection of loblolly pine appears to be reduced. Lower P. tinctorius infections coincide with lower levels of sucrose in the roots. On the other hand, Lamb and Richards (3) found that greater mycorrhizal infection of Monterey pine (Pinus radiata D. Don) and slash pine Pinus elliottii Engelm. on Australian soils occurred when P was applied. The objective of this study was to examine the relationship between nutrient levels and *P. tinctorius* ectomycorrhizal development on container-grown loblolly pine seedlings.

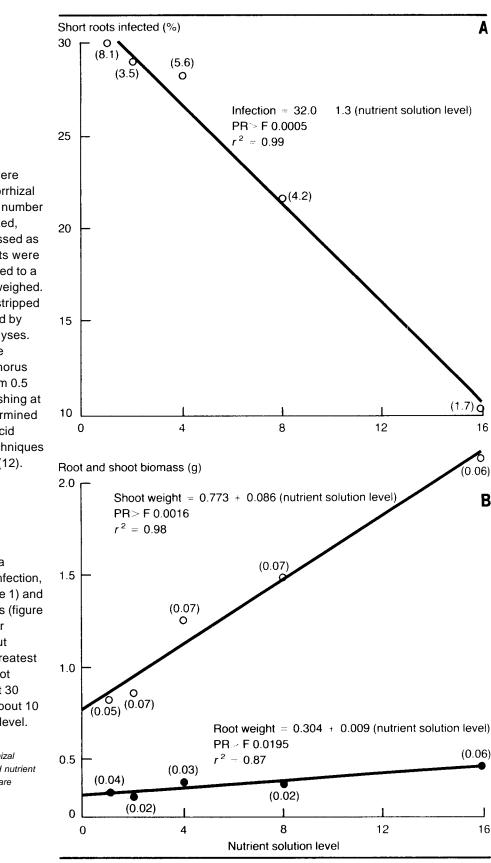
#### **Materials and Methods**

*P. tinctorius* fruiting bodies were collected in the fall from a 20-year-old loblolly pine (*Pinus taeda* L.) stand in Buchanan County, VA. Basidiospores were harvested, suspended in water, and added to vermiculite and peat (1 :1, v/v) that was being mixed in a cement mixer. Spores were applied at a rate equivalent to 250 grams per hectare. Spencer-Lemaire Rootrainers (Hillson model, 150 cubic centimeter per cavity) were filled with this mixture and two loblolly pine seeds of Virginia Piedmont origin were placed in each container. After germination, seedlings were thinned to one per container.

Four replications (eight seedlings per replication) of five nutrient solution levels (treatments) were used in this study (table 1). The five treatments consisted of a serial dilution of reagent grade chemicals. Molar ratios of 1 :1.2:1.1 :0.3 for KN0<sub>3</sub>, NH<sub>4</sub>S0<sub>4</sub>, H<sub>3</sub>PO<sub>4</sub>, and Fe (as Greenol), respectively, were used to provide the various amounts of nutrients. A serial dilution was used to establish treatment levels so that the most concentrated treatment was 16 times as concentrated as the lowest fertilization rate (table 1). Nutrients were added in 15 milliliters of water every 2 weeks. Additional watering occurred on a regular basis.

# Table 1—Amount of nitrogen,phosphorus, potassium, and ironadded every 2 weeks during the20-week study period

Amount of nutrient (mg)				
Solution level	Ν	Ρ	ĸ	Fe
1 ×	0.25	0.16	0.21	0.09
<b>2</b> ×	0.50	0.32	0.42	0.18
<b>4</b> ×	1.00	0.64	0.84	0.36
<b>8</b> ×	2.00	1.28	1.68	0.72
16×	4.00	2.56	3.36	1.48



After 20 weeks, seedlings were harvested. The degree of mycorrhizal infection was calculated as the number of individual short roots colonized. divided by the total, and expressed as a percentage. Roots and shoots were separated at the root collar, dried to a constant weight at 65 °C, and weighed. After weighing, needles were stripped from the stems and composited by replicate for foliar N and P analyses. Foliar N was determined by the Kjeldahl technique (2). Phosphorus was extracted with 6 N HCI from 0.5 gram of tissue, following dry-ashing at 450 °C. Phosphorus was determined by the Murphy-Riley ascorbic acid technique (13). Regression techniques were used to analyze the data (12).

#### **Results and Discussion**

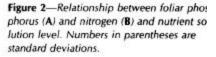
Fertilizer concentration had a significant effect on degree of infection, shoot weight, root weight (figure 1) and on foliar N and P concentrations (figure 2). The highest rates of fertilizer resulted in larger seedlings, but mycorrhizal colonization was greatest at the lower rates. Maximum root system colonization was about 30 percent, which decreased to about 10 percent at the highest fertilizer level.

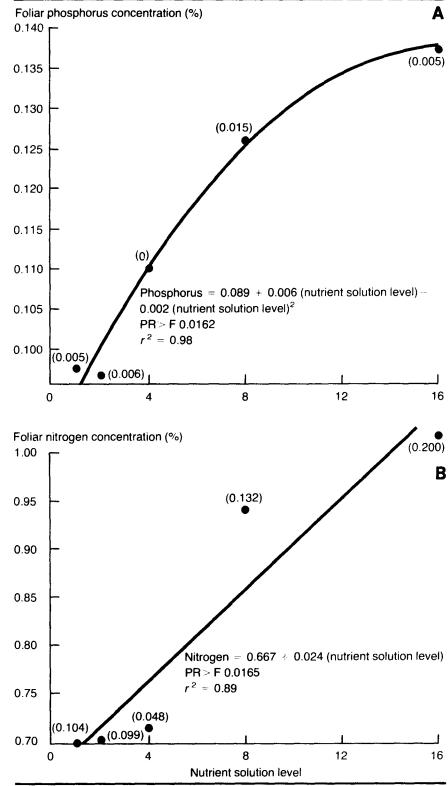
**Figure 1**-Relationship between mycorrhizal infection (**A**), seedling biomass (**B**), and nutrient solution level. Numbers in parentheses are standard deviations.

Foliar nutrient levels increased with fertilizer rate (figure 2). In the most concentrated treatment, foliar N and P levels were 1.02 and 0.14 percent, respectively. Although the N/P ratio (1.56 milligrams per milligram) in the nutrient solution was greater than the N/P ratio (1.21) used by other researchers (9, 10), there seemed to be insufficient N even at the highest treatment level. Nitrogen is generally considered deficient when foliar levels are below 1.2 percent, whereas 0.1 percent is a commonly accepted critical level for P (14).

There were also significant relationships between foliar nutrients and seedling biomass and infection (figures 3 and 4). Shoot weight increased but root infection decreased with increasing foliar N and P levels. Seedling size continued to increase as foliar P levels rose above 0.1 percent. This can be explained by the fact that N was limiting but P was available in sufficient quantities. Additional increments of N and P then resulted in luxury consumption of P, while seedling growth was still responding to added N.

Although root weight was used as a measure of root size, an estimate of total root length or surface area would have been more meaningful. There were distinct morphological differences observed



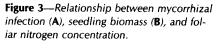


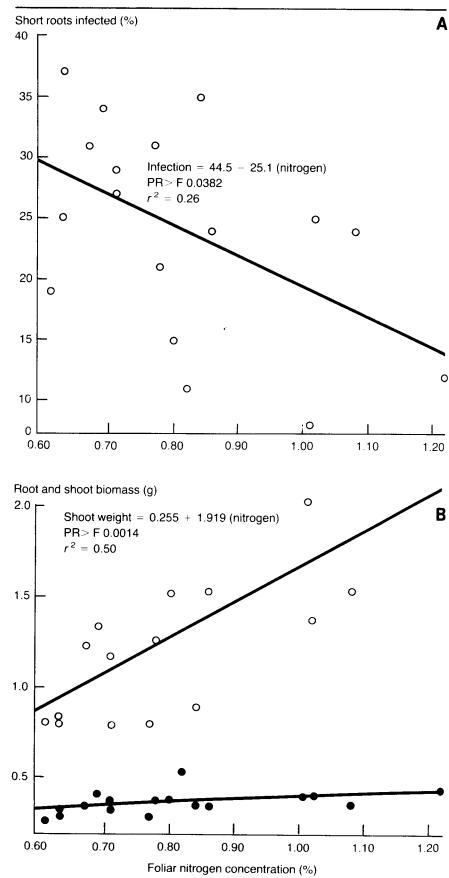
between root systems at different nutrient levels. Root systems of seedlings receiving the highest treatment were short and stocky. Although they weighed less, the root systems subjected to lower nutrient levels were longer, more fibrous, and had a greater degree of branching.

In order for seedlings to survive in harsh environments, they need an adequate root system to absorb moisture and nutrients. Seedlings receiving the greatest amount of nutrients in this study, although larger, could be at a disadvantage when outplanted. They would require greater amounts of moisture to meet the demands of their larger foliar biomass, but their root sys tems had a smaller surface area and less absorptive capacity. The likelihood that the smaller trees receiving the lowest levels of nutrients would be more successfully established is increased by the greater degree of mycorrhizal infection.

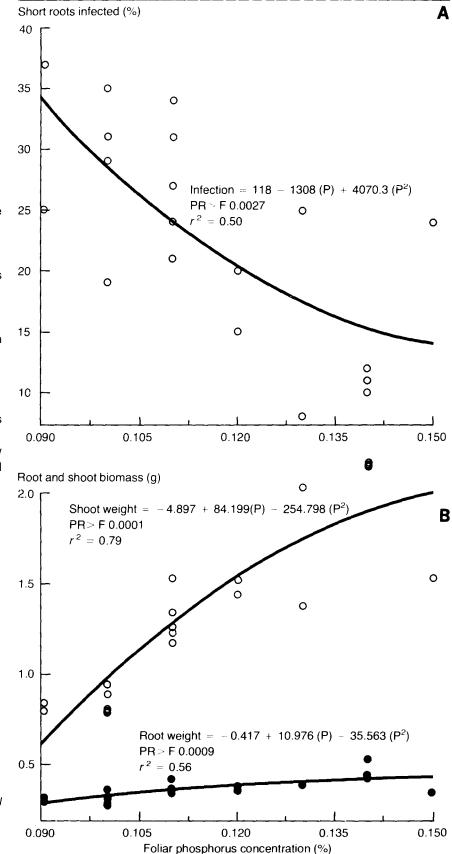
## Conclusion

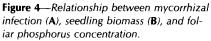
Although nutrients are necessary to develop healthy containerized seedlings, it appears that there is an optimal level above which additional nutrient supplies produce seedlings that may be more sensitive to drought and less able to





adapt to stress. Seedling size, root system morphology, and mycorrhizal colonization need to be optimized. An ideal fertilization program might be one that allows the seedlings to attain just enough N and P to reach foliar levels considered adequate for good growth. At higher levels, mycorrhizal infection is reduced and root systems undergo morphological changes resulting in smaller surface areas. In this study, level 4 x (1.0 milligram N, 0.6 milligram P, and 0.8 milligram K) may represent the best level of nutrient supplementation because infection is still high and seedlings are significantly larger than those receiving lower levels of nutrients. Ultimately, seedlings should be outplanted to determine how these morphological and physiological properties affect establishment and early growth.





### Literature Cited

- Bowen, G.D. Mineral nutrition of ectomycorrhizae. In: Marks, G.C.; Koslowski, T.T., eds. Ectomycorrhizae: Their ecology and physiology. New York: Academic Press; 1973; 444 p.
- Bremner, J.M. Total nitrogen. In: Black, ed. Methods of soil analysis. Madison, WI: American Society of Agronomy; 1965:1149-1178.
- Lamb, R.J.; Richards, B.N. Inoculation of pines with mycorrhizal fungi in natural soils. II. Effects of density and time of application of inoculum and phosphorus amendment on seedling yield. Soil Biology and Biochemistry 6:173-177: 1974.
- Marx, D.H. Use of specific mycorrhizal fungi on tree roots for forestation of disturbed lands. In: Proceedings, Conference on Forestation of Disturbed Surface Areas, 1976 April 14-15; Birmingham, AL. U.S. Department of Agriculture, Forest Service, State and Private Forestry; International Forest Seed Co. 1976: 47-65.

- Marx, D.H. Synthesis of ectomycorrhizae on loblolly pine seedlings with basidiospores of *Pisolithis tinctorius*. Forest Science 2:13-20; 1976.
- Marx, D.H.; Artman, J.D. *Pisolithis tinctorius* ectomycorrhizae improve survival and 11. growth of pine seedlings on acid coal spoils in Kentucky and Virginia. Reclamation Review 2:2331; 1979. 12.
- Marx, D.H.; Hatch, A.B.; Mendicino, J. F. High soil fertility decreases sucrose content 13. and susceptibility of loblolly pine roots to ectomycorrhizal infection by *Pisolithus tinctorius*. Canadian Journal of Botany 55:15691574; 1977.
- Marx, D.H.; Ross, E.W. Aseptic synthesis 14. of ectomycorrhizae on *Pinus taeda* by *Thelophora terrestris*. Canadian Journal of Botany 48:197-198; 1969.
- Ruehle, J.L. Inoculation of containerized loblolly pine seedlings with basidiospores of *Pisolithis tinctorius*. Res. Note SE-291. Asheville, NC: U.S. Department of Agriculture, Forest Service; 1980. 4 p.

- Ruehle, J.L.; Marx, D.H. Developing ectomycorrhizae on containerized pine seedlings. Res. Note SE-242. Asheville, NC: U.S. Department of Agriculture, Forest Service; 1977; 8 p.
  - Ruehle, J.L.; Marx, D.H. Fiber, food, fuel, and fungal symbionts. Science 206:419-122; 1979.
- 12. SAS Institute. SAS 79.5. Raleigh, NC: SAS Institute; 1979; 494 p.
  - Watanabe, F.S.; Olsen, S.R. Test of an ascorbic acid method for determining phosphorus in water and NaHCO<sub>3</sub> extracts from soil. Soil Science Society of America, Proceedings 29:677-678; 1965.
  - Wells, C.G.; Crutchfield, D.M.;
    Berenyl, N.M.; Davey, C.B. Soil and foliar guidelines for phosphorus fertilization of loblolly pine. Res. Pap. SE110. Asheville, NC: U.S. Department of Agriculture, Forest Service: 1973.
    15 p.