

PROLIFERATION OF SHORT SHOOTS IN CONTAINER-GROWN SCOTS PINE SEEDLINGS: A MEANS TOWARD RAPID VEGETATIVE MULTIPLICATION

Richard H. Hillson and Bruce P. Dancik

Department of Forest Science
University of Alberta
Edmonton, Alberta, Canada

Short shoots (also referred to as needle bundles, dwarf shoots, brachyblasts, fascicles, and spur shoots) provide a partial solution to a twofold problem:

1. Rootability in pine cuttings is highest when the stock plant is in the seedling stage, and decreases rapidly with age.
2. Young seedlings, by virtue of their size, yield only a limited number of cuttings.

Short shoots from young pines are not difficult to root but seldom develop terminal buds, and eventually die (4); however, it has long been known that terminal (interfoliar) bud development on short shoots follows damage to the branch terminal bud (5), (1), (10), (3). Isikawa and Kusaka (8) decapitated leaders of 12-year-old Japanese black pines and obtained short shoots with predeveloped interfoliar buds. These were subsequently rooted and developed into normal, upright trees. The need for development of an interfoliar bud while the short shoot is still on the tree has also been recognized by Rudolph and Nienstaedt (9) and by Hoff and MacDonald (7).

More recently Hillson (6) reported that interfoliar shoots exhibiting high rootability were produced on 5-year-old Scots pines (*Pinus sylvestris*) subjected to a 24-hour photoperiod following removal of all leader and branch apices. Attention was drawn to the

Rapid proliferation of short shoots followed removal of terminal bud clusters on 9-month-old container-grown Scots pine (*Pinus sylvestris*) seedlings. Growth was stimulated by continuous light providing 2,000 fc intensity at average plant height.

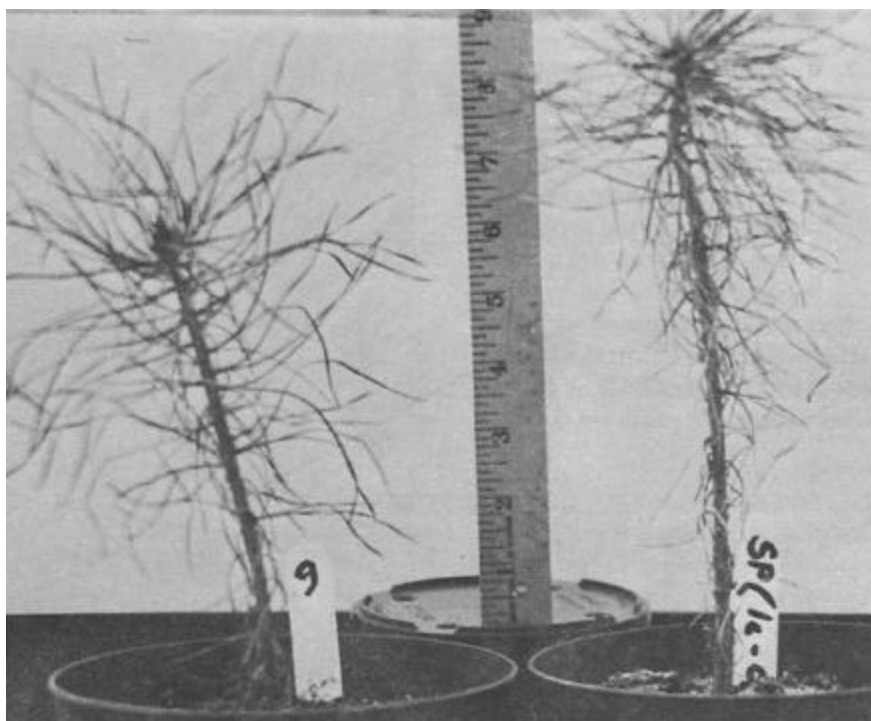


Figure 1.—Before terminal buds were removed on October 11, seedlings averaged approximately 8 in. in height.

suggestions of Dickson and Thomson that the proliferating short shoot may represent a reversion to the juvenile phase, and the report suggested that scion wood from mature trees be grafted onto seedling stock. Successful grafts exposed to extended photoperiods, following decapitation of leaders and branches, might then produce juvenile (rootable) interfoliar shoots and thus overcome the age problem.

Whitehill and Schwabe (11) showed that outgrowth of interfoliar shoots could be

influenced by precutting treatment of the stock plant with cytokinins and other growth substances. The number of shoots was increased (at the expense of size) by the cytokinin treatment and was depressed by the gibberellin treatment. These authors also reported success in inducing rootable interfoliar shoots on mature scion wood grafted onto juvenile rootstocks.

Three facts are thus apparent:

1. Interfoliar shoots can be used to increase the number of rootable cuttings available from young (and therefore

small) rootstocks.

2. Interfoliar shoots must be induced while the short shoot is still on the parent tree.
3. Interfoliar shoots induced on scions from mature trees can help overcome the problems of age-influenced decrease in rootability.

This note is concerned with the early production of interfoliar shoots on pine seedlings as a first step in obtaining rootable cuttings.

Method

As part of a continuing study, terminal bud clusters were removed on October 11 from five 31-week-old container-grown Scots pine seedlings. The seedlings were then placed in a growth-chamber providing a light intensity of 2000 fc at average-plant height. Temperature was held within the 15°-21° C range throughout the trial. These seedlings, obtained from the Alberta Provincial Tree Nursery, were produced in an intensive-culture programme involving an initial (winter) greenhouse phase and a secondary (summer) outdoor phase. On October 11 they averaged approximately 8 in. in height (fig. 1), and each seedling bore a number of short shoots (in the axils of primary leaves) on the upper half of the single, unbranched stem (fig. 2).

Results

By October 19, only 8 days after the seedlings were placed in the growth chamber, one to several

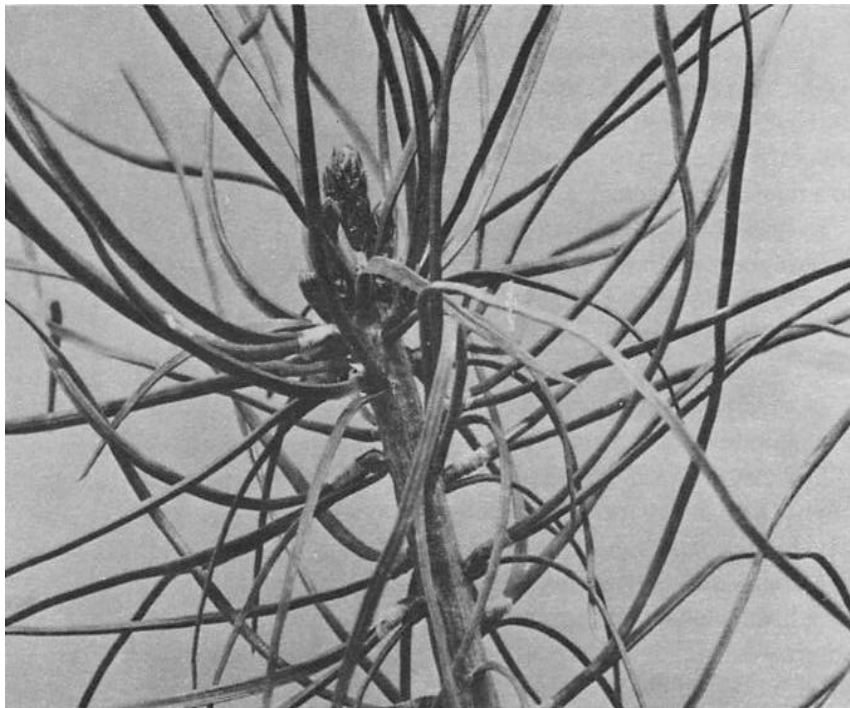


Figure 2.—Each seedling bore a number of short shoots (in the axils of primary leaves) on the upper half of the single, unbranched stem.

interfoliar buds were macroscopically visible on all five seedlings. These developed rapidly into actively growing interfoliar shoots – without any intervening period of apparent quiescence – and, by November 4, some shoots were over an inch in length. Bud development was mainly basipetal, but on one seedling the lower short shoots proliferated earlier and more vigorously than the upper.

By November 7, short-shoot primordia were visible in the axils of scale leaves borne on some of the interfoliar shoots. The scale leaves themselves were much

modified, being long, stiff, and much flattened. Because of this, young interfoliar shoots resembled small pineapple crowns.

Interfoliar shoots were measured on November 15, 35 days after the beginning of the trial (table 1).

Discussion

The method outlined above would be most appropriate for use in conjunction with intensive-culture, container-growing programmes, such as those now operated by Provincial, State, and Federal departments, especially where these are associated with studies involving such areas as

disease resistance and site evaluation. The multiple ramets produced would allow for replicated testing of many infection levels or on many sites.

Use of the method for multiplication of selected genetic material for outplanting depends on the effectiveness of other studies aimed at determining criteria for correlating juvenile characteristics with adult performance. As yet these are not considered to be fully reliable.

It is also interesting to conjecture that, if the proliferated short shoot really does represent a reversion to the juvenile phase, then the possibility exists for the multiplication of mature trees selected in the field or in test plantations. This could be accomplished by grafting selected scion wood onto seedling stock and by inducing proliferation of short shoots on successful grafts (6), (11). This might overcome the age-related decrease in growth rate which has been observed in conventional grafts, and in rooted cuttings from mature trees (2).

Finally, it is worth pointing out that, once the parent material is available, the procedure outlined in this note has the following advantages:

1. It requires only the simplest manipulation of the stock plants.
2. It requires little in the way of equipment other than some means of providing a suitable

Table 1.—*Number and length of interfoliar shoots 35 days after the beginning of the trial*

Tree No.	Interfoliar shoots (Number)	Average length of interfoliar shoots (mm)	Range (mm)
1	10	38	16-64
2	8	27	5-69
3	10	28	14-42
4	8	28	6-50
5	5	22	5-31

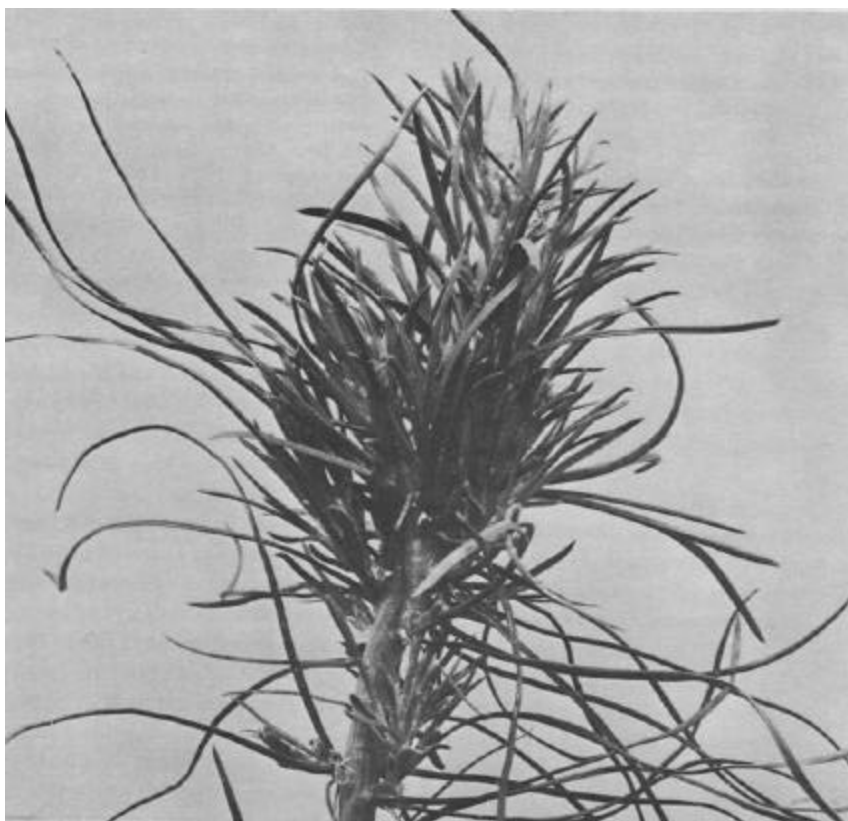


Figure 3.—*Interfoliar buds developed rapidly into interfoliar shoots, some over an inch in length by November 4.*

light source and an acceptable ambient temperature. The standard tree-nursery greenhouse would be adequate.

3. It does not require the application of growth-regulating chemicals.

Literature Cited

1. Borthwick, A. W.
1899. On interfoliar buds in pines. *Trans. and Proc. Bot. Soc. Edinburgh* 21:154-158.
2. Brix, H. and R. van den Driessche.
1977. Use of rooted cuttings in reforestation. *British Columbia For. Serv., Can. For. Serv. Joint Rep. No. 6.*
3. Cooperrider, C. K.
1938. Recovery of ponderosa pine reproduction following injury to young growth. *Plant Physiol.* 13:5-27.
4. Delisle, A. L.
1942. Histological and anatomical changes induced by indoleacetic acid in rooting cuttings of *Pinus strobus* L. *Va. J. Sci.* 3:119-124.
5. Dickson, A.
1885. On the development of bifoliar spurs into ordinary buds in *Pinus silvestris*. *Trans. and Proc. Bot. Soc. Edinburgh* 16:258-261.
6. Hillson, R. H.
1970. Some aspects of the vegetative propagation of pines by means of cuttings and detached short shoots. Unpubl. M.Sc. rep. submitted at Wye College (University of London), Ashford, Kent.
7. Hoff, R. J. and G. I. McDonald.
1968. Rooting of needle fascicles from western white pine seedlings. *For. Serv. U.S. Dep. Agric., Intermountain Forest and Range Exp. Sta. Res. Note INT-80.* 6 p.
8. Isikawa, H. and M. Kusaka.
1959. The vegetative propagation of cuttings of *Pinus* species I. Vegetative propagation of Japanese black pine using leaf bundles. *Bull. For. Exp. Sta. Meguro, Tokyo.* 116:50-64.
9. Rudolph, T. D. and H. Nienstaedt.
1964. Rooting, shoot development and flowering of jack pine needle fascicles. *Silv. Genet.* 13:118-123.
10. Thomson, R. B.
1914. The spur shoot of the pines. *Bot. Gaz.* 57:362-385.
11. Whitehill, S. J. and W. W. Schwabe.
1975. Vegetative propagation of *Pinus sylvestris*. *Physiol. Plant.* 35:66-71.