Interactions Between Seedbed Mulches and Seedling Disease Development¹

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Mulching of seedbeds immediately after seed sowing has been and remains a common practice in the production of bare root seedlings in forest tree nurseries. A good mulch provides an important cover for newly sown seeds, enhances seedbed soil stability. facilitates seed germination via maintenance /retention of adequate soil moisture, and buffers seed and seedlings from extremes in soil surface temperatures. Organic mulches provide the added benefit of building or maintaining desirable levels of soil organic matter, which in turn enhance soil structure, water and nutrient relations, and soil microbiological interactions (Adams 1966. Bollen and Glennie 1961. Davey and Krause 1980, Mannering and Meyer 1963, Van Nierop and White 1958).

¹ Paper presented at the Northeastern and Intermountain Forest and Conservation Nursery Association Meeting, St. Louis, Missouri, August 2-5, 1993

² Respectively: Forest Pathologist, Florida Division of Forestry, FDACS, Gainesville, FL; Plant Pathologist, U.S.D.A. Forest Service, Olustee, FL; Forestry Supervisor II, Florida Division of Forestry, FDACS, Gainesville, FL Although not commonly perceived as a contributor to disease development or a control for nursery seedling diseases, mulches may, depending upon a variety of factors, be either. In this paper we

- highlight reports from the literature where mulches were known or believed to have influenced seedling diseases (esp. in forest tree nurseries)
- 2 allude to mechanisms involved in mulch-disease interactions
- 3 consider criteria for mulch selection with respect to disease relations, and
- 4 review a case history from a Florida forest nursery where mulching has been employed successfully for control of *Rhizoctonia* blight of longleaf pine (*Pinus palustris* Mill.).

Abstract – Seedbed mulches might be thought to have little influence on disease relationships in forest tree nurseries. In fact, mulches may serve as sources of pathogenic inoculum, or provide conditions that either favor or prevent the development of seedling diseases. This paper highlights aspects of mulchdisease interactions and summarizes a Florida study in which mulching provided useful control of *Rhizoctonia* seedling blight of longleaf pine (*Pinus palustris*). A basic understanding of the biology of soilborne pathogenic and beneficial microorganisms and the influence that cultural practices such as mulching may have on diseas e development is both needed and encouraged.

INVOLVEMENT OF MULCHES IN NURSERY DISEASES: A LITERATURE PERSPECTIVE

Mulches as sources of inoculum.

The type and source of available mulching materials and their potential for carrying pathogenic organisms are issues that need to be considered when selecting a mulch for nursery seedbeds. Certain materials have a higher probability of being contaminated with pathogenic organisms than others (Bloomberg 1963,1985, Hoitink et al. 1976, Schönhar 1968). For instance, the fungus Sphaeropsis sapinea (Fr). Dyko & Sutton can infect and sporulate on cones, needles, and branches of Ponderosa pine (*Pinus ponderosa* Dougl. ex Laws) and red pine (Pinus resinosa Ait.) (Sinclair et al. 1987), and one should avoid use of these materials as mulches where susceptible species are grown (Peterson and Nichols 1989). Sand and peat moss can carry pathogenic species of Pythium and Fusarium (Sutherland 1984), and warnings have been issued to Southern nursery managers about the potential for

pine needles to carry *Fusarium* spp. and the charcoal root rot fungus, *Macrophomina phaseolina* (Tassi) Goid. (Cordell and Lantz, U.S.D.A. Forest Service, FIDM Letter to Southeastern Forest Nurserymen, February 29, 1980). Needles of some conifer species may carry needle cast fungi such as *Meria laricis* Vaill. and *Lophodermium* spp. and should be avoided where diseases caused by such fungi pose a hazard to seedling crops (Sutherland 1984, Staley and Nichols 1989).

Influence of mulches on disease development

In addition to the possibility of introducing pathogenic fungi into nurseries, certain types of organic matter may differentially favor the development of pathogenic

and beneficial fungus populations. For example, Bloomberg (1963) found that species of Trichoderma predominated on hemlock sawdust, but species of Alternaria were favored by straw. *Trichoderma* spp. are generally believed to restrict the growth of pathogenic spp., but Alternaria spp. are regarded as pathogens in some situations. Table 1 provides an overview of some mulchdisease interactions on various seedling crops as reported in the literature. Note that the influence of mulches can be variable. sometimes promoting and sometimes preventing disease development. Mechanisms involved in such influences are myriad, often subtle, complex and may be biological, chemical, and/or physical in nature (Table 2).

Table 2. Some ways in which mulches may influence disease development.		
Biologically	promote/suppress pathogen activity stimulate competi- tive/antagonistic microflora	
Chemically	pH alterations nutrient relations direct phytotoxicity pesticidal (suppres - sion)	
Physically	soil stability temperature H ₂ O aeration flotation	

Soil moisture and temperature relationships

Mulches influence soil temperature and soil moisture. Such

Table 1. Some examples of mulch-disease interactions on various seedling crops.

Mulch Material	Сгор	Effect	Ref.	
Sawdust	Longleaf Pine	Decreased D.O. ¹ (Rhizoctonia)	Davis 1941	
Sphagnum		Decreased D.O.	Winters 1949	
Coir Dust	Mahogany	Decreased D.O.	Barnard 1950	
Doug-fir Sawdust	Strawberries	Increased Phytophthora	Vaughan et al. 1954	
Various	Ginko	Decreased M. phaseolina	Fang et al. 1956	
Sawdust/Wood Chips		Increased Nematodes	Van Nierop & White 1958	
"Dark" Sawdust	Douglas -Fir	Increased Heat Lesions	Salisbury & Long 1959	
Pine Sawdust/Bark	Pinus sp.	Decreased D.O.	Vaartaja & Bumbieris 1965	
Sawdust	Ginger	Decreased Root-Knot Nematodes	Colbran 1974	
Redwood Bark/Sawdust	Red Fir & Douglas -Fir	Decreased Phoma Blight	Kliejunas et al. 1985	
Hardwood Bark/Wood Chips	Longleaf Pine	Decreased Rhizoctonia	Gilly et al. 1985	

¹D.O. = damping-off

Table 3. Some factors to consider when selecting mulching materials for forest nursery seedlings.

Availability	Cost
Timing	Rate
Material	natural/synthetic
	organic/inorganic
Age/Condition	fresh/aged/
	composted
	chemistry (phyto-
	toxicity, C:N ratio)
Color	temperature
	relations
Texture	stability
	H ₂ O relations
	aeration
Source/	contaminant/
Handling	pathogenic organ-
	isms
	beneficial/microor-
	ganisms
I	

influences can indirectly influence the development of biotic diseases, either positively or negatively, depending on particular host-pathogen relationships. For example, Vaughan et al. (1954) found that increased soil moisture and reduced soil temperatures associated with the use of Douglas-fir sawdust mulch increased red-stele disease in strawberries caused by the water mold fungus. Phytophthora fragariae Hickman. In contrast, the use of mulches has been credited with the control of charcoal root rot on ginkgo (Ginko *biloba L.*) seedlings (Fang *et al.*) 1956). In this instance, the use of a mulch on nursery beds maintained soil temperatures below those considered optimal (c. $32^{\circ}C$) for infection by the pathogen, Macrophomina phaseolina.

Alternatively, the influence of mulches on soil and/or soil surface temperatures can directly affect seedlings by causing or preventing abiotic injuries or

"diseases". For example, dark colored mulches may absorb solar heat, and thereby increase the chances of heat injury (Barnard 1990, Boyce 1961, Peace, 1962). The use of light colored mulches, on the other hand, can reduce heat injury by reflecting insolation and dispersing heat. However, under certain circumstances, light colored or reflective mulches also may contribute to heat injury of seedlings. Richards (1970) found that the use of fiberglass and spruce needles as mulches could greatly increase temperatures immediately above seedbeds and that these elevated temperatures were damaging to white spruce [*Picea glauca* (Moench) Voss] and Norway spruce [P. abies (L.) Karst.].

Physical relationships

The ability of a mulch to act as a barrier against wind and rain and hold a soil in place can play a significant role in the prevention of some diseases. Kliejunas et al. (1985) reported that mulch composed of redwood bark and sawdust reduced soil splash, soil cone formation and the incidence of Phoma blight caused by Phoma eupvrena Sacc. in fir and Douglasfir seedbeds in a California nursery. Soil splash that normally occurred during winter months led to soil cone formation around seedlings. This soil cover was believed to reduce seedling vigor and provide an environment favorable to disease development. The use of a mulch in Florida nurseries has produced a similar beneficial effect with regard to the development of Rhizoctonia blight of longleaf pine (see Case History below).

Chemical relationships

Chemicals that are naturally associated with certain types of organic matter (...mulches) can suppress growth and development of soilborne pathogens. Extracts from sawdust of Pinus banksiana Lamb., Populus spp. (Carlson and Belcher 1970), and hardwood bark compost (Kai 1990) can inhibit the growth of certain species of root-infecting fungi. Spencer and Bensen (1982) found that extracts of pine bark suppressed the growth of Phytophthora cinnamomi Rands. but noted that the suppressive effects of extracts from hardwood bark compost were stronger and more consistent.

The exact mechanisms by which chemical exudates from organic matter affect specific soilborne fungi are not always clear. In some instances, it appears that the leachates can inhibit the vegetative growth of fungi (Huang and Kuhlman 1991), but in other cases, the chemical extracts may interrupt stages in the life cycle of the fungus. For instance, Hoitink et al. (1977) found that leachates from composted hardwood bark could inhibit sporangium formation and lyse germ tubes of zoospores of P. *cinnamomi*, thus preventing host infection. Zoospore production in another water mold, Pythium aphanidermatum (Edson) Fitz. has been shown to be inhibited by extracts from pine bark (Huang and Kuhlman 1990).

Water soluble extracts that are leached from various types of mulches can be highly beneficial to seedlings when they suppress fungal pathogens. However, caution is advised in the selection

and handling of organic mulches because some organic materials may be phytotoxic. Organic materials that receive insufficient oxygen during storage or composting can undergo anaerobic breakdown and the resulting fermentation products may be toxic to seedlings (Bollen and Lu 1970, Hoitink and Fahey 1986). Reindeer-moss has been reported to be phytotoxic to jack pine (P. banksiana) and white spruce [Picea glauca (Moench) Voss] (Fisher 1979), and grain straw residues proved to have deleterious effects on black spruce seedlings [P. mariana (Mill.) B.S.P.1 (Jobidon et al. 1989) in instances where these materials were used as mulches. In both cases, phytotoxicity was apparently indirect via disruption of mineral nutrient uptake (P in the former case, and Mn in the later case). Fisher (1979) postulated that the reduction in P uptake may have resulted from an alteration of mycorrhizal symbiosis.

Biological relationships

Other processes through which mulches may be beneficial in the prevention of seedling diseases involve numerous mechanisms which may be collectively referred to as biological control. Populations and activity of soilborne pathogens are often suppressed by the action of competitive, antagonistic, or hyperparasitic microorganisms which may be stimulated by either the food base or environmental niche provided by certain organic materials and/or mulches. A detailed treatise of specific mechanisms involved in these often complex processes is well beyond the scope of this paper. Accordingly, interested readers are referred to any number of excellent overviews (Adams 1990, Baker 1987, Baker 1968, Baker and Snyder 1970, Baker and Cook 1974, Boland 1990, Campbell 1989, Cook and Baker 1983, James *et al.* 1992).

USE OF MULCHES TO CONTROL *RHIZOCTONIA* BLIGHT OF LONGLEAF PINE: A FLORIDA CASE HISTORY

In the southeastern U.S. longleaf pine (Pinus palustris) seedling crops are frequently damaged by Rhizoctonia spp. which appear to opportunistically infect seedlings impacted by "sand splash" (Davis 1941, English and Barnard 1989, English et al. 1986). Davis (1941) reported reduced damage when seedling crops were mulched with sawdust. Faced with unacceptable losses in the late 1970's and early 1980's at the Florida Division of Forestry's Andrews Nursery, Gilly et al. (1985) initiated a field trial to evaluate the influence of cultural practices (i.e., seedling density and mulching materials) and fungicidal sprays on the growth and development of longleaf pine seedlings and the incidence and severity of Rhizoctonia blight in longleaf pine seedbeds. Results of the fungicide trials were inconsistent, whereas mulching with hardwood (bark and wood) chips provided consistent reductions in the occurrence of disease with no detrimental effects on seedling production. In the following paragraphs, we review aspects of the mulching

Table 4. Mulches tested for effects on the development of Rhizoctonia Blight of longleaf pinea			
Material	Rate		
Pine straw (1X)	Operationally		
	applied (6-12 mm		
	layer)		
Pine straw (2X)	2X operational rate		
	(12-25 mm layer)		
Hardwood bark/			
wood chips	12-25 mm layer		
Hydromulch®	1180 kg/h ^a		
^a All applied immediately after seeding.			

trial and present a summary and discussion of pertinent results.

Materials & Methods

Seedbeds were prepared according to customary procedures which included soil fumigation with methyl bromide (98% active ingredient @ 393 kg/ha) and a pre-plant incorporation of 15-0-15 fertilizer at 225 kg/ha). Seeds were sown at densities of 350 seeds/m2 (high) and 150seeds/m2 (low). Four mulch treatments (Table 4) were applied in each of four replicate plots (ca. 60 X 1.25 m each) distributed in a randomized complete block fashion within each of the two seedbed planting densities.

Systematically located 0.37 m² life history plots (1 per test plot) were utilized to monitor treatment effects on seed germination and seedling survival. The number of seeds (or seedlings) in each plot was recorded

- 1 immediately following seed sowing,
- 2 34 days later, and
- 3 at the end of the growing season.

Mulch Treatment	Germination %	Seedlings/m3	Total Oven Dry Weight (g)	Root Collar Diameter (mm)
	Low Densit	y Planting (150 seed	l/m²)	
Hydromulch™	26.2a	30.5a	8.6a	9.2a
Hardwood chips	54.7c	86.5b	7.6a	8.6a
1X pine straw	40.2b	76.5b	8.3a	8.8a
2X pine straw	58.3c	82.4b	8.4a	8.8a
	High Densit	y Planting (350 seed	l/m²)	
Hydromulch™	31.1a	90.1	5.6a	7.4a
Hardwood chips	53.3c	164.c	4.9a	7.3a
1X pine straw	42.2b	125.b	5.4a	7.3a
2X pine straw	54.4c	158.c	5.4a	7.1a

Table 5. Production parameters as affected by planting density and mulch treatment ^a

^a Values within columns and seedbed planting densities followed by same letter are not significantly different at $P \le 0.05$. Oven dry weight and RCD's differed significantly ($P \le 0.05$) between low and high density plantings.

All test plots were carefully examined for evidence of *Rhizoctonia* blight on each of six dates between July 5 and November 11. Wooden markers were placed in the ground beside each diseased seedling, and at the end of the test periods, the total number of markers in each infection center (i.e., 1 or more seedlings showing symptoms of infection) were recorded. Incidence and severity of *Rhizoctonia* blight were evaluated by

- 1 the number of *Rhizoctonia* infection centers/10 m of bed length,
- 2 the average number of infected seedlings/infection center,
- 3 the total number of infected seedlings/10 m bed length, and
- 4 average percentage seedling loss.

At the end of the nursery growing season, two 0.37 m²

subplots were systematically established in each density-mulch plot. The number of seedlings in each of these subplots were counted and ten seedlings were randomly selected from each subplot for determinations of a) root collar diameters (RCD) and b) oven-dry weights.

Results and Discussion

Differences in germination between mulch treatments were significant (Table 5). Highest germination occurred in beds mulched with hardwood chips and 2X pine straw, followed by 1X pine straw and Hydromulch®. This order was consistent in both planting densities. Germination in Hydromulch[®] plots was so low that end-of-season seedbed stocking (i.e., seedlings per unit area) in "high planting density" Hydromulch® plots was comparable to that in the "low planting density" plots of the other three mulches. Not surprisingly, seedlings grown in sparsely stocked Hydromulch® plots were somewhat larger than seedlings grown in plots with other mulches. Within each planting density, seedlings from beds mulched with hardwood chips were slightly smaller than seedlings from 1X or 2X pine straw plots, even though seedbed densities were comparable. These differences, however, were not statistically significant. Within all mulch treatments, seedlings attained greater oven-dry weights and RCD's when grown at the lower planting density.

Table 6 summarizes results with respect to disease incidence and severity within treatment plots, based on observations performed at the end of the growing season. In both seedbed planting densities the highest losses to disease were sustained in plots mulched with pine straw at the 1X rate. In contrast, losses in plots mulched with hardwood chips were lowest among all four

Mulch Treatment	No. Inf. Centers/ 10m Bed ^a	Mean No. Seedlings/ Inf. Center a	No. Dead Seedlings/ 10m Bed	Seedling Loss (%)	
Low Density Planting (150 seed/m ²)					
Hydromulch™	6.6a	1.5a	9.9	2.7	
Hardwood chips	3.3a	1.6a	5.3	0.5	
1X pine straw	6.8a	5.3a	36.0	3.9	
2X pine straw	4.5a	6.7a	30.2	3.0	
	High Dens	ity Planting (350 seed	1/m²)		
Hydromulch™	7.1 ac	5.5a	39.1	3.5	
Hardwood chips	5.5a	6.4a	35.2	1.7	
1X pine straw	11.6bc	14.8a	171.7	11.3	
2X pine straw	5.1a	10.6a	54.1	2.8	

Table 6. Effects of planting density and mulches on Rhizoctonia Blight of longleaf pine.

^a Values within columns and seedbed planting densities followed by same letter do not significantly differ at $P \le 0.05$.

treatments. Losses indicated for Hydromulch®-treated plots are deceptively low and misleading due to the very poor germination/stand survival which occurred in this treatment (Table 5). Considering both production parameters (Table 5) and disease occurrence data (Table 6), it is clear that the hardwood chip mulch provided the best treatment overall.

Davis (1941) reported reduced incidence of *Rhizoctonia* infections when longleaf pine seedbeds were mulched with 6-12 mm of sawdust and suggested that mulching might provide useful disease control. Our results support this contention, and of the materials we tested, hardwood chips are clearly the material of choice. While we cannot, on the basis of our studies, eliminate the possibility of some type of biological control being active here, it appears likely that simple physical or mechanical control of sand splash is clearly involved. In this regard, our findings parallel those of Kliejunas *et al.* (1985).

CONCLUSIONS

Results of the case study (above), together with the results and observations reported by others, should encourage forest nursery managers to learn all they can regarding the biology of pathogens causing problems in their respective nurseries. In some instances, the control of diseases caused by pathogenic organisms may be effected via alteration of cultural practices such as seedbed mulching. In Florida, nursery managers have learned that even "re-mulching " longleaf pine seedbeds (i.e. applying a second application of mulch to seedbeds, months after seed germination) is beneficial with respect to reducing losses to *Rhizoctonia* spp.

(unpublished data and observations).

One must keep in mind, however, that nature and events are not always predictable, and that even the best of techniques can have an "Achilles heal". In one Florida situation. Rhizoctonia caused a foliage blight of loblolly pine (P. taeda L.) because the material used for mulch on seedbeds floated up to and lodged in the needles of seedlings during heavy rains, thus facilitating the transfer of inoculum from the soil surface to susceptible needles. Although damage was minor and primarily restricted to low-lying areas of seedbeds, the situation was nonetheless disconcerting for the nursery manager.

A basic understanding of pathogen biology and disease epidemiology (development) coupled with a keen sense of observation and a willingness to experiment are advocated as useful tools in the nursery managers constant battle to develop and maintain healthy seedling crops.

LITERATURE CITED

- Adams, J. E.1966. Influence of mulches on runoff, erosion and soil moisture depletion. Soil Sci. Soc. Amer. Proc. 30: 110-114.
- Adams, P. B., 1990. The potential of mycoparasites for biological control of plant diseases. Ann. Rev. Phytopathol. 28: 59-72.
- Baker, K. F. 1987. Evolving concepts of biological control of plant pathogens. Ann Rev. Phytopathol. 25: 67-85.
- Baker, K. F., and Cook, R. J. 1974. Biological control of plant pathogens. Freeman. San Francisco. 433 p.
- Baker, K. F., and Snyder, W. C. eds. 1970. Ecology of soil-borne plant pathogens: prelude to biological control. Univ. Calif. Press. Berkeley, CA 571 p.
- Baker, 8.1968. Mechanisms of biological control of soil-borne plant pathogens. Ann. Rev. Phytopathol. 6: 263-294.
- Barnard, E. L. 1990. Groundline heat lesions on tree seedlings.Fl. Dept. Agric. & Consumer Serv., Div. Plant Ind. Pathol. Circ. No. 338. 2 p.
- Barnard, R. C. 1950. Coir dust mulch in nursery practice. Malaysian Forester 13: 165.
- Bloomberg, W. J.1963. Use of organic residues in forest nurseries. For. Ent. Path. Branch, Can. Dept. For., Bi-m. Progress Report 19:4 pp.

Bloomberg, W. J. 1985. The epidemiology of forest nursery diseases. Ann. Rev. Phytopathol. 23: 83-96.

- Boland, G. J.1990. Biological control of plant disease with fungal antagonists: challenges and opportunities. Can. J. Plant Pathol.12: 295-299.
- Bollen, W. B., and Glennie, D. W. 1961. Sawdust, bark, and other wood wastes for soil conditioning and mulching. For. Prod. Journal 11: 38-46.
- Bollen, W. B. and K. C. Lu. 1970. Sour sawdust and bark - its origins, properties, and effect on plants. USDA For. Serv., Res. Pap. PNW-108:13 pp.
- Boyce, J. S. 1961. Forest pathology. McGraw-Hill Book Co. New York. 572 p.
- Campbell, 8.1989. Biological control of microbial plant pathogens. Cambridge Univ. Press Cambridge. 218 p.
- Carlson, L. W. and J. Belcher. 1970. Effect of nursery soil and soil amendment extracts on growth of damping-off fungi and conifer seed germination. For. Ent. Path. Branch, Can. Dept. For., Bi-m. Res. Notes 26:26 pp.
- Colbran, R. C. 1974. Nematode control in ginger with nematocides, selection of planting material, and sawdust mulch. Queensl. J. Agric. Anim. Sci. 31: 231-235.
- Cook, R. J., and Baker, K. F. 1983. The nature and practice of biological control of plant pathogens. Amer. Phytopathol. Soc. St. Paul, MN. 539 p.
- Davey, C. B., and Krause, H. H. 1980. Function and maintenance of organic matter in forest nursery soils. Pages 130-165 in: Proceeding of the North American Forest Tree Soils Workshop. Syracuse, NY, July 28-August 1, 1980. U.S.D.A. Forest Service, Canadian

Forestry Service and S.U.N.Y Syracuse. 333 p.

- Davis. W. C. 1941. Damping-off of longleaf pine. Phytopathology 31:1011-1016.
- English, J. T., and Barnard, E. L. 1989. *Rhizoctonia blight* of longleaf pine. Pages 66-67 in: Forest Nursery Pests. C. E. Cordell *et al.* Tech. Coords. U.S.D.A. For. Serv. Agric. Handbook No. 680.184 p.
- English, J. T., Ploetz, R. C., and Barnard, E. L. 1986. Seedling blight of longleaf pine caused by a binucleate *Rhizoctonia* solani-like fungus. Plant Dis. 70: 148-150.
- Fang, C. T., Yien, S. Y., Lee, C. T., and Wang, K. M. 1956. Experiments on the control of stem rot disease of ginko caused by Macrophomina phaseoli. Acta Phytopathologica Sinica 2(1): 43-54.
- Fisher, R. F. 1979. Possible allelopathic effects of reindeer-moss (*Cladonia*) on jack pine and white spruce. For. Sci. 25: 256-260.
- Gilly, S. P., Barnard, E. L., and Schroeder, R. A. 1985. Field trials for control of *Rhizoctonia* blight of longleaf pine seedlings: Effects of seedbed planting densities, fungicides, and mulches. Pages 476-485 in: Proceedings of the International Symposium on Nursery Management Practices for the Southern Pines. D. B. South, ed. Alabama Agric: Stn. Auburn Univ. and International Union of Forest Research Organizations. 594 pp.
- Hoitink, H.A.J., Herr, L. J., and Schmitthener, A. F. 1976.Survival of some plant pathogens during composting of

hardwood tree bark. Phytopathology 66: 1369-1372.

- Hoitink, H.A.J., Van Doren, D.M., Jr., and Schmitthener, A. F. 1977. Suppression of *Phytophthora cinnamomi* in a composted hardwood bark potting medium. Phytopathology 67: 561-565.
- Hoitink, H. A. J. and P. C. Fahy. 1986. Basis for the control of soilborne plant pathogens with composts. Ann. Rev. Phytopathol. 24:93-114.
- Huang, J. W. and E. G. Kuhlman. 1991. Mechanisms inhibiting damping-off pathogens of slash pine seedlings with a formulated soil amendment. Phytopathology 81:171-177.
- James, R. L., Dumroese, R. K., and Wenny, D. L. 1992. Principles and potential for biocontrol of diseases in forest and conservation nurseries. Pages 122-131 in: Proceedings, Western Forest Nursery Assoc. T. D. Landis, Tech. Co-ord. U.S.D.A. For. Serv. GTR RM-221. 151 p.
- Jobidon, R. Thibault, J. R., and Fortin, J. A. 1989. Effects of straw residues on black spruce seedling growth and mineral nutrition under greenhouse conditions. Can. J. For. Res. 19: 1291-1293.
- Kai, H., Veda, T., and Sakaguchi, M. 1990. Antimicrobial activity of bark-compost extracts. Soil Biol. Biochem. 22:983-986.
- Kliejunas, J. T., Allison, J. R., McCain, A. H., and Smith, R. S., Jr. 1985. Phoma blight of fir and Douglas-fir seedlings at a California nursery. Plant Dis. 69:773-775.
- Mannering, J. V. and Meyer, L. D. 1963. The effects of various rates of surface mulch on

infiltration and erosion. Soil Sci. Soc. Amer. Proc. 27: 84-86.

- Peace, T. R. 1962. Pathology of trees and shrubs. Oxford University Press. 753 p.
- Peterson, G. W. and T. H.
 Nicholls. 1989. Diplodia Blight.
 In Forest Nursery Pests. C. E.
 Cordell, R. L. Anderson, W. H.
 Hoffard, T. D. Landis, R. S.
 Smith, and H. V. Toko, editors.
 USDA Forest Service, Washington, D. C., pp. 31-33.
- Richards, N. A. 1970. Adverse effects of mulching Spruce seedlings. Tree Planters' Notes 21:11-12.
- Schonhar, S. 1968. The occurrence of damping-off fungi in compost. Allg. Forst. Jagdztg. 139:227-229.
- Sinclair, W. A., H. H. Lyon, and W. T. Johnson. 1987. Diseases of Trees and Scrubs. Cornell University Press, Ithaca. 574 PP.
- Spencer, S., and Benson, D. M. 1982. Pine bark, hardwood bark compost, and peat amendment effects on development of Phytophthora spp. and lupine root rot. Phytopathology 72: 346-351.
- Staley, J. M. and T. H. Nicholls. 1989. Lophodermium Needle
 Cast. In Forest Nursery Pests.
 C. E. Cordell, R. L. Anderson,
 W. H. Hoffard, T. D. Landis, R.
 S. Smith, Jr., and H. V. Toko,
 editors. USDA Forest Service,
 Washington, D. C. pp. 49-51.
- Sutherland, J. R. 1984. Pest management in Northwest bareroot nurseries. In Forest Nursery Manual: Production of Bareroot Seedlings. M. L. Duryea and T. D. Landis, editors. Martinus Nijhoff/Dr. D. W. Junk Publishers, The Hague, Boston, Lancaster. pp. 203-210.

- Vaartaja, O., and Bumbieris, M. 1965. Control of conifer damping-off in South Australia. Plant Dis. Rep. 49: 504-506.
- Van Nierop, E. T., and White, D. P. 1958. Evaluation of several organic mulching materials on a sandy loam forest nursery soil. J. For. 56: 23-27.
- Vaughan, E. K., Roberts, A. N., and Mellenthin, W. M. 1954. The influence of Douglas-fir sawdust and certain fertilizer elements on the incidence of red-stele disease of strawberry. Phytopathology 44: 601-603.
- Winters, H. F. 1949. Seedbed treatments. Rep. Fed. Exp. Sta. Puerto Rico. pp. 7-8.