

From Forest Nursery Notes, Winter 2013

199. Intraspecific variation in *Pinus pinaster* PSII photochemical efficiency in response to winter stress and freezing temperatures. Corcuera, L., Gil-Pelegrin, E., and Notivol, E. PLoS One 6(1):e28772. 2011.

Intraspecific Variation in *Pinus Pinaster* PSII Photochemical Efficiency in Response to Winter Stress and Freezing Temperatures

Leyre Corcuera*, Eustaquio Gil-Pelegrin, Eduardo Notivol

Forest Resources Unit, Research and Technology Agricultural Centre (CITA), Government of Aragon, Zaragoza, Spain

Abstract

As part of a program to select maritime pine (*Pinus pinaster* Ait.) genotypes for resistance to low winter temperatures, we examined variation in photosystem II activity by chlorophyll fluorescence. Populations and families within populations from contrasting climates were tested during two consecutive winters through two progeny trials, one located at a continental and xeric site and one at a mesic site with Atlantic influence. We also obtained the LT_{50} , or the temperature that causes 50% damage, by controlled freezing and the subsequent analysis of chlorophyll fluorescence in needles and stems that were collected from populations at the continental trial site. *P. pinaster* showed sensitivity to winter stress at the continental site, during the colder winter. The combination of low temperatures, high solar irradiation and low precipitation caused sustained decreases in maximal photochemical efficiency (F_v/F_m), quantum yield of non-cyclic electron transport (Φ_{PSII}) and photochemical quenching (qP). The variation in photochemical parameters was larger among families than among populations, and population differences appeared only under the harshest conditions at the continental site. As expected, the environmental effects (*winter* and *site*) on the photochemical parameters were much larger than the genotypic effects (*population* or *family*). LT_{50} was closely related to the minimum winter temperatures of the population's range. The dark-adapted F_v/F_m ratio discriminated clearly between interior and coastal populations. In conclusion, variations in F_v/F_m , Φ_{PSII} , qP and non-photochemical quenching (NPQ) in response to winter stress were primarily due to the differences between the winter conditions and the sites and secondarily due to the differences among families and their interactions with the environment. Populations from continental climates showed higher frost tolerance (LT_{50}) than coastal populations that typically experience mild winters. Therefore, LT_{50} , as estimated by F_v/F_m , is a reliable indicator of frost tolerance among *P. pinaster* populations.

Citation: Corcuera L, Gil-Pelegrin E, Notivol E (2011) Intraspecific Variation in *Pinus Pinaster* PSII Photochemical Efficiency in Response to Winter Stress and Freezing Temperatures. PLoS ONE 6(12): e28772. doi:10.1371/journal.pone.0028772

Editor: Steven M. Theg, University of California – Davis, United States of America

Received: August 11, 2011; **Accepted:** November 15, 2011; **Published:** December 29, 2011

Copyright: © 2011 Corcuera et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This research was sponsored by the Spanish INIA (Instituto Nacional de Investigación y Tecnología Agraria y Alimentaria) project RTA2005-00131-00-00 and MCI (Ministerio de Ciencia e Innovación) project AGL-2010-21153-CO2-02. The funders had no role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: lcorcuera@aragon.es

Introduction

Pinus pinaster progeny trials have been established throughout Spain for conservation purposes and to analyze the growth and physiological adaptations of different seed sources to several soil and meteorological conditions [1]. These trials were designed with a nested structure to examine families within populations. The term “family” refers to a group of individuals that have one or both parents in common (half-sib and full-sib families, respectively). “Population” denotes the group of individuals within which there is gene exchange, and “provenance” refers to the geographic origin of the population. Natural populations have been subjected to selection by their particular set of local environmental conditions and may differ in performance when grown at a common site. Progeny trials are the best way to evaluate the genetic value of selected parents to determine the population best suited to particular climatic conditions. The relationships among the traits that are related to yield (survival, wood volume) and wood quality (polycyclism) have allowed the selection of populations that are well suited to the environmental conditions

at several trial sites [2]. In particular, the populations and families evaluated here have shown differentiation and plasticity in growth and physiological parameters related to drought, such as carbon isotope composition [3], vulnerability to xylem embolism [4] and accumulation of phytohormones [5].

As in other evergreen species, the leaves of the maritime pine allow CO_2 uptake during the whole year, whenever favorable climate conditions occur. Physiological parameters related to CO_2 fixation, such as the rate of photosynthesis and the activity of photosystem II (PSII), decline with abiotic stress [6]. Drought stress is the main factor that restrains growth and survival in the Mediterranean climate. In humid and cold forest zones, low temperatures are the limiting factor. However, frost-induced reductions in photosynthesis are reversible. The recovery time after frost varies with species, frost intensity, cold hardiness [7] and the light exposure of the needles [8]. Natural populations experience several simultaneous environmental stresses that have interactive effects. The combination of low winter temperatures and high solar radiation increases the amount of absorbed light that cannot be used by the plant, leading to greater reductions in

PSII photochemical efficiency in conifers [9] and broadleaf evergreens [10]. In Mediterranean areas with a continental climate (large seasonal temperature differences: hot summers and cold winters), cold hardiness and frost resistance of evergreens are a matter of concern [11]. In this case, winter stress can be more harmful for the photosynthetic apparatus than summer stress [12] as low winter temperatures coincide with high solar irradiation and drought stress. However, the photoprotection processes, such as the increase of non-photochemical quenching (NPQ), ensure the dissipation of excess light and prevent chronic photoinhibition, allowing the recovery of photosynthesis and PSII photochemical efficiency when temperatures increase in the spring [13].

Chlorophyll fluorescence has been a useful tool for allowing plant physiologists to detect the effects of biotic and abiotic stresses on light-processing physiology [14]. Changes in fluorescence parameters in response to environmental stresses, such as low [e.g., 15] and high temperatures [e.g., 16], high light intensity [e.g., 17] and drought [e.g., 18], have been widely documented at the species level. Intra-specific variation in the tolerance to low temperatures among populations is essential for seed transfer between geographic regions during reforestation [19,20]. A positive relationship between the latitude of provenance and F_v/F_m ratio after frost events was observed in *Pinus contorta* and *Pinus sylvestris* populations [21]. At the family level, Koechn et al. [22] found genetic variation in qP and F_v/F_m and a positive relationship between qP and growth in *Pinus elliotii*.

Chlorophyll fluorescence is also an accurate indicator of freezing injury. Freeze tolerance, or the ability of plants to survive subfreezing temperatures, can be measured by the dark-adapted F_v/F_m ratio. However, chlorophyll fluorescence as a screening method for freezing injury in conifers has been investigated mainly at the species level. It has been shown as a fast and reliable technique for establishing the freeze temperature that causes 50% damage (LT_{50}) in *Pinus sylvestris* needles and stems (cortical bark chlorenchyma) [23]. The estimation of the LT_{50} of different genotypes by the F_v/F_m ratio has been investigated mainly in crop plants [e.g., 24]. Few studies have measured intraspecific variation in LT_{50} [25] and cold tolerance [26] by means of chlorophyll fluorescence in forest tree species.

To date, only the short-term and seasonal responses of photosynthesis to temperature have been documented in *P. pinaster* populations [27]. Meanwhile, to our knowledge, no studies have investigated the effect of winter stresses (high irradiation, low temperatures and drought) or the temperature that causes freezing injury by means of chlorophyll fluorescence at the species, population or family level in *P. pinaster*. In this work, we tested several *Pinus pinaster* genotypes with different field survival rates in provenance-progeny trials during two consecutive winters at two sites that vary in productivity due to contrasting altitudes and precipitation regimes. One site is at a low elevation near the Atlantic and is exposed to wet and mild winters, and the other site is at an interior mid-high elevation with colder and drier winters. Our objectives were (1) to assess the environmental and genetic variability in PSII photochemical performance in response to winter stress (by means of Φ_{PSII} , F_v/F_m , qP and NPQ) and (2) to determine whether the LT_{50} (estimated by F_v/F_m) was related to the original climate of the populations. Our results indicate that there is genetic variation in photochemistry in response to winter stress, mainly at a family level. We present evidence that population differences in the freezing tolerance of needles and stems are consistent with the winter temperatures of the climate of origin, providing evidence in support of local adaptation.

Results

The effect of climate on photochemical efficiency and survival

The monthly means of the minimum autumn and winter temperatures were lower in 2005–2006 than in 2006–2007 at both sites (Fig. 1A). The study was conducted during the late winter of 2006 (cold winter) and 2007 (warm winter, Fig. 1A). Two progeny trials were conducted (Fig. 2). The total autumn and winter precipitation at the mesic site (ME) was four to five times higher than that at the xeric site (XE). The cumulative radiation at the XE was close to two (winter 2005–2006) or three times higher (winter 2006–2007, Fig. 1B) than that at the ME.

The first component of the principal component analysis (PCA) absorbed 99% of the total variation. The main variables were the following, in order of relevance: continentality (extreme winter and summer temperatures), annual precipitation, precipitation during the wettest quarter and precipitation during the coldest quarter. Populations were arranged according to the bioclimatic index (Table 1): *Arenas* and *Tamrabta*, the populations from continental climates, followed by *Oria* from south Spain, are subjected to more stressful conditions at their sites of origin, similar to those experienced at the xeric site. The *Mimizan* population, from France, is native to a climate similar to that at the mesic site.

Despite the low autumn and winter precipitation at the XE, no trees at any of the trial sites had signs of water stress. Predawn water potential was lower after the cold winter (ME: -0.03 ± 0.01 MPa in the cold winter; -0.01 ± 0.01 MPa in the warm winter; XE: -0.87 ± 0.03 MPa in the cold winter; -0.58 ± 0.03 MPa in the warm winter).

As expected, environmental conditions (*winter*, *site*, and their interaction, *site x winter*) had a major influence on photochemical parameters (Table 2). All three populations were sensitive to the combination of winter stresses during the cold winter at the continental site, which was characterized by low temperatures, low precipitation (approximately 200 mm in autumn and winter) and high irradiation (PAR above 2000 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$).

After the cold winter, there was a reduction in F_v/F_m values, i.e., a down-regulation of PSII efficiency due to a cumulative effect of winter stresses at the XE (F_v/F_m : $0.70\text{--}0.73 \pm 0.01$ compared to $0.83\text{--}0.85 \pm 0.006$ at the ME, Fig. 3A1). F_v/F_m values were within [28] or below [29] the ranges described in previous studies of *Pinus halepensis* and did not decrease significantly, indicating that the remaining PSII reaction centers were photochemically active during the winter.

The proportion of energy used for photochemistry (i.e., the actual photochemical efficiency, Φ_{PSII}) and the proportion of PSII open centers, qP , had similar behavior at both sites and during both winters (Figs. 3B1, 3C1, 3B2 and 3C2). Both Φ_{PSII} and qP exhibited maximum values after the mild winter and minimum values after the cold winter at both sites.

At the xeric trial, the French population (*Mimizan*) was more sensitive to drought than any other population. In 2005, survival rates were 48%, 50% and 40% for the *Arenas*, *Oria* and *Mimizan* populations, respectively. In 2006, survival rates were 35%, 36% and 23% for the *Arenas*, *Oria* and *Mimizan* populations, respectively. High mortality was observed during both years due to severe drought and degraded soils. In the mesic trial, survival was close to 100%, and the posterior mortality (<5%) was the consequence of human error, which occurred when several of the trees were cut with farming implements while performing mechanical weed control.

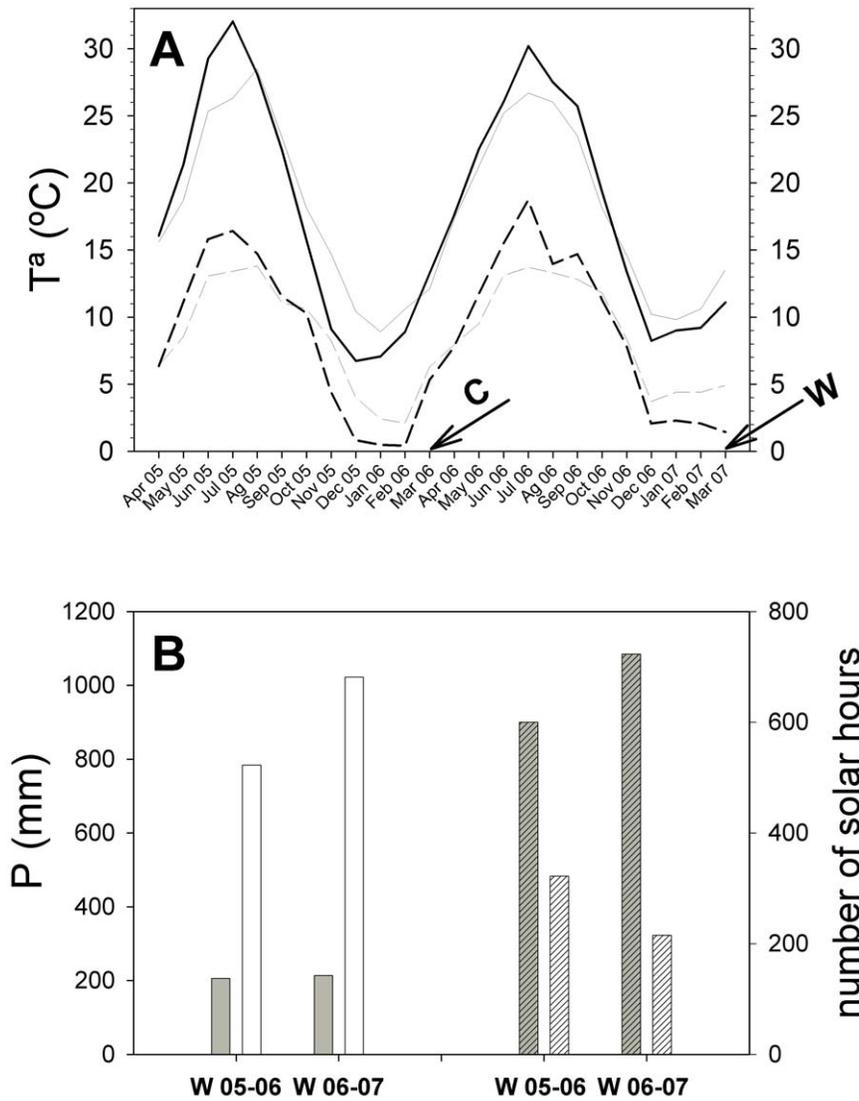


Figure 1. Climatic data. (A) Monthly mean of the maximum temperatures in XE (thick line) and ME (hairline) and of the minimum temperatures in XE (dashed line) and ME (dashed hairline). Arrows indicate the measurement dates. C: cold winter, W: warm winter. (B) Sum of the precipitation in autumn and winter in XE (grey bars) and ME (white bars). Sum of the solar hours in XE (gray hatched bars) and ME (white hatched bars). W 05–06: winter of 2005–2006, cold winter. W 06–07: winter of 2006–2007, warm winter. doi:10.1371/journal.pone.0028772.g001

Intraspecific winter variation in PSII photochemical parameters

Differences among populations were only observed at the XE (Table 3), and some significant heritabilities were found at the XE as well (Table 4). Population differences in Φ_{PSII} , qP and NPQ after the warm winter (Table 3; Figs. 3B2, 3C2, 3D2) and in NPQ after the cold winter were observed (Table 3; Fig. 3D1). The *Mimizan* population showed a significantly higher Φ_{PSII} and qP after the warm winter than the *Arenas* and *Oria* populations (Fig. 3B2, 3C2; Table 3). Heat dissipation was lower in the *Arenas* population, which inhabits a cold climate, after both winters (Figs. 3D1, 3D2; Table 3).

The variation in photochemical parameters among families is shown in Figs. 4 and 5. Differences among the families were significant for Φ_{PSII} , qP and NPQ (Table 2) and were significant for F_v/F_m when the parameters were obtained for each site and winter (Table 3). The family and its interaction with *winter*, *site* and *site x winter* were all significant sources of variance for all of the parameters (Table 2).

Survival rates were not related to the photochemical performance of the populations. At the XE, the *Mimizan* population exhibited lower survival rates that were not associated with an inferior photochemical performance.

Intraspecific variation in LT_{50}

Figure 6 shows the evolution of F_v/F_m in needles (A) and stems (cortical bark chlorenchyma, B) exposed to frost temperatures. There was a clear differentiation between the interior and coastal populations. Needle tolerance to freezing temperatures was higher in the populations from locations with a continental climate and low winter temperatures, *Arenas* and *Tamrabta* (LT_{50} : -28.7 ± 0.40 and -28.0 ± 0.62 , respectively, $R^2 = 0.99$, $P < 0.0001$; Fig. 6A, Table 1). The populations from locations with mild winters, *Oria* and *Mimizan*, exhibited lower absolute values of LT_{50} (-23.5 ± 0.10 and -24.4 ± 0.31 , respectively, $R^2 = 0.99$, $P < 0.0001$; Fig. 6A). However, the frost resistance of the cortical bark chlorenchyma was only significantly higher in *Tamrabta*

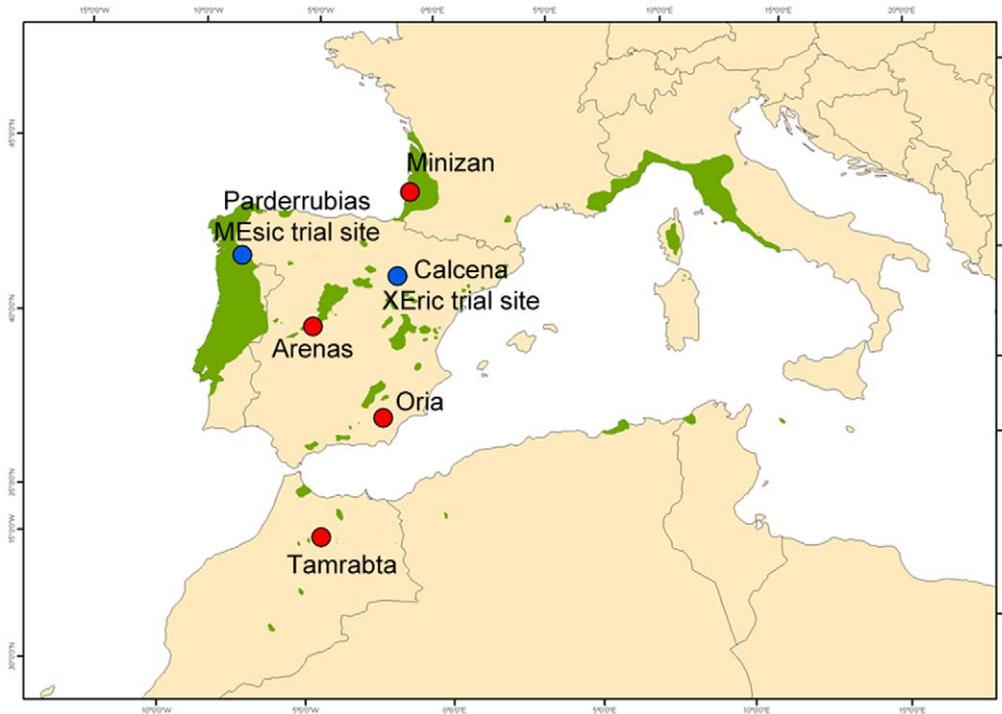


Figure 2. Locations of the populations and trial sites.
doi:10.1371/journal.pone.0028772.g002

(-35.4 ± 0.76 ; $R^2 = 0.99$, $P < 0.0001$) than in the rest of the populations (*Arenas*: -31.1 ± 1.62 , *Oria*: -29.1 ± 0.88 and *Mimizan*: -30.0 ± 1.30 ; $R^2 = 0.99$, $P < 0.0001$; Fig. 6B). Population differences in LT_{50} were correlated with the mean minimum temperature of the coldest month at the seed source (Fig. 7). LT_{50} decreased with decreasing mean minimum temperatures of the seed source for both needles ($R^2 = 0.85$, $P < 0.05$) and stems ($R^2 = 0.84$, $P < 0.05$). The ranking of populations according to LT_{50} was similar for needles and stems.

Relationship among the photochemical parameters

High values of qP were associated with high values of Φ_{PSII} (Table 5) in general, and when they were separated by *site* and *winter* (Table 6). Similarly, high values of F_v/F_m were correlated

with high values of NPQ (Table 5), in general and when they were separated by *site* and *winter* (Table 6). F_v/F_m was positively correlated with Φ_{PSII} and qP at both sites after the cold winter. At the *XE*, NPQ was negatively correlated with Φ_{PSII} after the warm winter and with qP after the cold winter (Table 6).

Discussion

The effect of winter stress on PSII photochemical parameters

Environmental factors (*site*, *winter*, *site x winter*) contributed to the phenotypic variation in photochemical parameters to a greater degree than did genetic sources of variation (*Pop*, *Fam*; Table 7 and 8). Most of the variation in photochemical parameters was

Table 1. Location and climatic data for the provenance-progeny trials and seed sources.

Locality	Calcena (XE)	Arenas	Tamrabta	Oria	Parderrubias (ME)	Mimizan
Bioclimatic Index	-794	-778	-613	-587	841	1026
Latitude (N)	41°37'	40°30'	33°71'	37°30'	42°14'	44°8'
Longitude (W)	1°44'	4°24'	4°74'	2°20'	7°56'	1°10'
Elevation (m)	1017	1359	1600	1232	460	0-80
P (mm)	461	692	950	351.5	722	1202
T (°C)	12.3	14.6	13.1	14.4	14.4	13.0
TM (°C)	28.6	34.2	33.0	30.0	27.8	25.0
Tm (°C)	1.1	0.3	-0.9	3.0	1.5	2.0

Populations and sites are arranged according to the Bioclimatic Index (lower values indicate drier continental climates). P: mean annual precipitation (mm); T: mean annual temperature (°C); TM: mean of the maxima in the month with the highest mean temperatures (°C); Tm: mean of the minima in the month with the lowest mean temperatures (°C); XE: continental and xeric site; ME: mesic site.
doi:10.1371/journal.pone.0028772.t001

Table 2. Summary of ANOVA significances.

Source of variation	Num DF	Den DF	F_v/F_m		Φ_{PSII}		qP		NPQ	
			F-Value	Pr>F	F-Value	Pr>F	F-Value	Pr>F	F-Value	Pr>F
Winter	2	541	111.96	<0.0001	101.11	<0.0001	185.55	<0.0001	45.10	<0.0001
Site	1	541	10.84	0.0011	2.45	0.1184	4.44	0.0355	3.47	0.0630
Winter x site	2	541	38.73	<0.0001	0.53	0.5906	3.81	0.0227	3.42	0.0334
Pop	2	541	0.55	0.5772	0.27	0.7657	1.03	0.3595	1.81	0.1654
Pop x winter	4	541	1.02	0.3975	1.79	0.1284	1.55	0.1851	2.46	0.0443
Pop x site	2	541	1.13	0.3225	1.56	0.2115	2.06	0.1290	6.11	0.0024
Pop (winter x site)	4	541	1.72	0.1448	3.28	0.0114	1.23	0.2968	1.02	0.3982
Fam (Pop)	42	541	1.33	0.0846	2.10	0.0001	4.10	<0.0001	1.58	0.0128
Fam (Pop x winter)	7	541	1.14	0.2028	1.77	0.0002	2.17	<0.0001	1.76	0.0002
Fam (Pop x site)	75	541	1.82	0.0807	1.34	0.2287	3.31	0.0019	3.15	0.0029
Fam (Pop x winter x site)	12	541	1.29	0.2176	1.78	0.0486	2.76	0.0012	2.08	0.0169

F_v/F_m : maximum potential PSII efficiency. Φ_{PSII} : actual PSII efficiency. qP : photochemical quenching. NPQ : non-photochemical quenching. Winter: studied period (winter 2005–06, winter 2006–07). Site: location of the provenance-progeny trials. Num DF: number of degrees of freedom. Den DF: Denominator of degrees of freedom. Pr: Probability. F: F-values. Bold numbers denote values for which Pr>0.05. doi:10.1371/journal.pone.0028772.t002

partitioned by these factors or was caused by family plastic responses to light, water and/or temperature stresses ($G \times E$ interactions, Table 7). This finding is consistent with Baquedano et al. [30]. The authors found that phenotypic plasticity blurred the ecotypic divergence of fluorescence traits, which could hide differences among *P. halepensis* populations and maintain genetic variation unavailable to selection.

The sensitivity of *P. pinaster* to low temperatures supports previous findings regarding Mediterranean species, e.g., low winter temperatures had a greater impact on F_v/F_m than high temperatures or drought during the summer in *Quercus suber* populations [31]. Moreover, when low winter temperatures were associated with low soil moisture, Mediterranean species showed the lowest values of F_v/F_m [32] and chronic photoinhibition [28]. Subtropical [33] and temperate [34] species have also displayed the lowest values of the maximum (F_v/F_m) and effective (Φ_{PSII}) quantum yield of PSII during winter.

Genetic variation in PSII photochemical parameters in response to winter stress and freezing temperatures (LT₅₀)

Genetic variation in *P. pinaster* photochemical parameters due to winter stress was found mostly at the family level (Table 8). There was greater intra-population variation than inter-population variation. Population variation was displayed exclusively at the xeric site. This finding is in line with a previous study [3] and suggests that population differentiation [35] and population selection [30] take place under adverse conditions. Measurements under favorable growing conditions in common garden experiments overcome the problem of confounding environmental effects with genotypic differences. However, some differences in physiological traits among genotypes that reflect adaptation to their climate of origin may be appreciated only when plants are exposed to stress. Population differences at the xeric site could be related to the higher cumulative radiation, lower precipitation and more extreme temperatures of the site (Fig. 2). This is consistent with the results of Colom et al. [36], who observed small but significant differences between populations in F_v/F_m and effective PSII quantum yield (Φ_{PSII}) at increasing light intensities. Aranda

et al. [31] also found differences among populations of *Quercus suber* during periods of low winter temperatures, when greater reductions in F_v/F_m (0.2–0.3) caused the highest population variance.

The low variation in the photochemical parameters attributed to the population effects found in this experiment is in agreement with Lopez et al. [37] and Baquedano et al. [30], who observed that photochemistry did not vary among populations in the control or drought treatments of *Pinus canariensis* and *Pinus halepensis*, respectively. Nevertheless, the small but highly significant population differences in photochemical performance that were expressed at the XE were related to the climatic origin of the seed source. The population from France, *Mimizan*, which experience mild winters due to the Atlantic influence, presented significantly higher values of Φ_{PSII} and qP during the warm winter. Photoprotection mechanisms, such as a high efficiency of heat dissipation, may have allowed *Mimizan* to reach higher Φ_{PSII} and qP values after the warm winter but not after the cold winter, when there were sustained decreases in F_v/F_m , Φ_{PSII} and qP . The *Arenas* population, originally from a location in interior and continental Spain with extreme maximum and minimum temperatures, displayed lower thermal energy dissipation (NPQ) in both winters.

Our results suggest that the LT₅₀ of needles and stems, estimated by F_v/F_m , can be used to test the freezing tolerance of *P. pinaster* intraspecifically. Differences among populations in needle LT₅₀ were consistent with the minimum temperatures of their climate of origin. This is in accordance with Climent et al. [38], who found that the LT₅₀, estimated by the electrolyte leakage method, was highly correlated with the mean temperature of the coldest month in Mediterranean pine species of contrasting thermal habitats. We observed less freezing damage in the *Tamraba* and *Arenas* populations, which are from continental climates, than in *Mimizan* and *Oria*, which are from coastal climates, reflecting adaptation to the ecological niches provided by the original climates. These results are consistent with similar studies in other coastal and interior conifer populations [26]. Even though the populations do not experience these minima temperatures at their sites of origin, this is useful information for possible reforestations in northern and colder latitudes in the future.

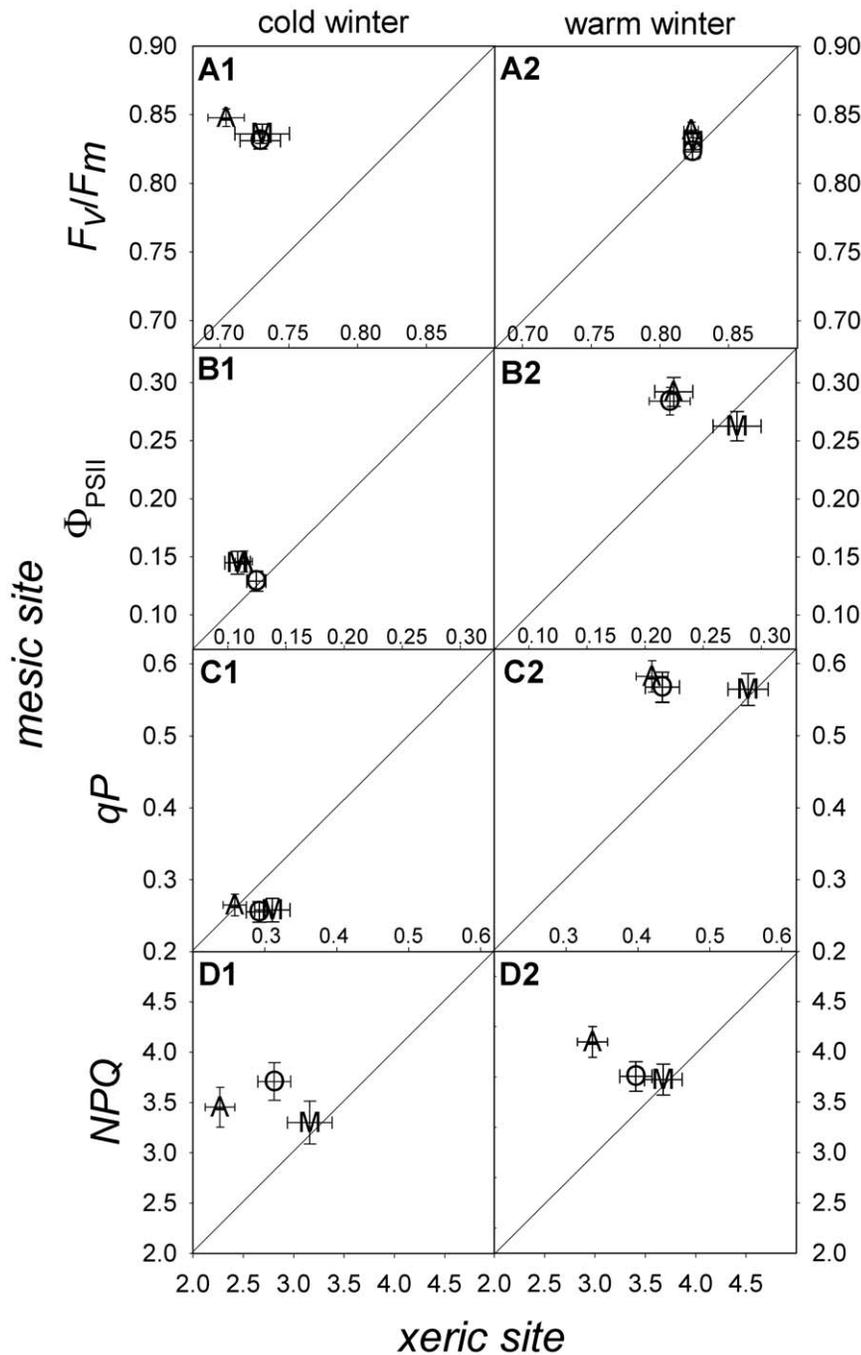


Figure 3. Population variation in the photochemical parameters by site and winter. Bi-directional least squared means \pm standard errors of the photochemical parameters for the populations (A: Arenas, O: Oria, M: Mizizan) present in the xeric (axis X) and mesic (axis Y) trial sites. Maximum potential (F_v/F_m) and actual (Φ_{PSII}) PSII efficiency and photochemical quenching (qP) and non-photochemical quenching (NPQ) after the cold (A1, B1, C1, D1) and warm winter (A2, B2, C2, D2) are shown. Diagonal lines indicate equal values at both sites. Values shown represent the mean of the values from 6 to 10 families from 3 populations. doi:10.1371/journal.pone.0028772.g003

The LT_{50} of the cortical bark chlorenchyma was more variable (higher standard errors) than needle LT_{50} . Plant material subjected to a range of freezing temperatures either escapes damage or is completely killed, and only a narrow range of temperatures induce partial damage. Taking into account the results of this research, we would expect that more accurate values would be obtained in future experiments that study several freezing temperatures in smaller increments ($1-2^\circ C$) within the

critical range of temperatures (from $-20^\circ C$ to $-30^\circ C$ for needles and from $-30^\circ C$ to $-40^\circ C$ for stems).

The maritime pine, as a Mediterranean conifer, must survive fluctuations in temperature, water soil availability, vapor deficits and high light irradiance during the year. The regulation of the electron transport processes by both environmental and genetic mechanisms is an advantage for the species. This study documents genetic variation in fluorescence traits, both between and within

Table 3. Summary of ANOVA significances by site and winter.

Winter	Site	Effect	Num DF	Den DF	F_v/F_m		Φ_{PSII}		qP		NPQ	
					F-Value	Pr>F	F-Value	Pr>F	F-Value	Pr>F	F-Value	Pr>F
Cold	XE	Pop	2	91	1.02	0.3632	0.83	0.4395	1.91	0.1547	6.47	0.0024
Cold	XE	Fam (Pop)	21	91	1.20	0.2694	2.45	0.0018	1.95	0.0159	1.88	0.0220
Cold	ME	Pop	2	62	1.80	0.1736	1.12	0.3320	0.11	0.8978	1.09	0.3433
Cold	ME	Fam (Pop)	19	62	1.98	0.0229	1.74	0.0532	1.31	0.2804	1.51	0.1142
Warm	XE	Pop	2	98	0.01	0.9895	2.77	0.0677	7.80	0.0007	4.66	0.0117
Warm	XE	Fam (Pop)	23	98	1.98	0.0113	2.13	0.0056	3.67	<0.0001	1.72	0.0358
Warm	ME	Pop	2	98	1.98	0.1442	1.50	0.2286	0.19	0.8265	1.85	0.1633
Warm	ME	Fam (Pop)	26	98	1.20	0.2562	2.59	0.0004	5.73	<0.0001	1.76	0.0251

Abbreviations are the same as those in Table 2.
doi:10.1371/journal.pone.0028772.t003

populations, but mostly within populations. However, variation due to environmental conditions accounted for the major proportion of the total variance. Although we found highly significant differences between families, most of the genetic variation was due to the interaction between family \times environment (site, winter, site \times winter). Further research is needed before considering intra-specific selection for photochemical efficiency in *P. pinaster*, and future studies should be directed toward finding differences among families. The investigation of a large number of genotypes over a wide range of environments in field tests would be necessary to understand the genes that regulate photosynthetic processes that can be monitored by chlorophyll fluorescence and their effects on survival and growth. The identification of quantitative relationships between abiotic stresses and photochemical activity could be useful to assess the plasticity of families and populations and for developing selection criteria for abiotic stress tolerance. In this work, we found that there is a potential for the selection of more frost tolerant *Pinus pinaster* populations, which is an advantage for a species that have to survive in unpredictable environments, especially in continental areas or at high altitudes.

Conclusion

We provide evidence for *P. pinaster* sensitivity to winter stress and for intraspecific variation in the PSII photochemical parameters in response to winter stress, mainly at the family level.

LT₅₀ obtained by F_v/F_m was consistent with the thermal ecological niche of the populations and can be reliably used to find differences in the frost tolerance of needles and stems among *P. pinaster* populations.

Table 4. Population heritabilities (h^2) with non-zero significance, by site and winter.

Winter	Site	F_v/F_m	Φ_{PSII}	qP	NPQ
Cold	XE	0	0	0	0.39
Cold	ME	0	0	0	0
Warm	XE	0	0.03	0.41	0.23
Warm	ME	0.06	0	0	0

Abbreviations are the same as those in Table 2.
doi:10.1371/journal.pone.0028772.t004

Materials and Methods

Ethics Statement

All necessary permits were obtained for the described field studies. Permissions required for field studies were obtained from the Environmental Departments of the Autonomous Governments of Aragon and Galicia.

Study site and plant material

The range of *Pinus pinaster* (Ait.) extends over the occidental Mediterranean basin and the southern European Atlantic coast of France and Spain. This relatively small area covers a wide range of climates, from arid to humid conditions, and altitudes from sea level up to 2000 m. It is a species widely used in Spanish reforestation programs and for tree breeding. Its highly fragmented distribution is explained by the discontinuity and high altitudes of the mountain ranges in southwestern Europe, which led to the isolation of geographically close populations and to several adaptations for growth and survival in distinct climates [39].

Open-pollinated siblings (individuals with one parent in common and the other parent unknown) were collected in natural stands of maritime pine in France, Spain and Morocco, and the seedlings were grown in nurseries. Progeny trials were established throughout Spain. We chose two progeny trials, located at Parderrubias, NW Spain (mesic site, ME) and Calcena, NE Spain (xeric site, XE). Sites were chosen to compare tree behavior under contrasting temperature and precipitation regimes. The ME is situated near the Atlantic Ocean, with a wetter and milder climate than the XE, which is continental with lower winter temperatures. Both sites undergo drought during the summer. Climatic data were obtained from the meteorological stations of Allariz, approximately 14 km from the ME (42°11'N, -7°48'W, 476 m a.s.l.), and Aranda de Moncayo, approximately 24 km from the XE (41°35'N, -1°47'W, 827 m a.s.l.).

At the ME, two-year-old seedlings were planted in 2005, separated from each other by 2 \times 3 m, in a randomized complete block design with 4 replications of 71 blocks, 225 families and 4 plants per experimental unit (a total of 16 plants per family). At the XE, one-year-old seedlings were planted in 2004, separated from each other by 2 \times 3 m, in an α -lattice incomplete block design with 3 replications of 65 blocks, 8 families per block and 4 plants per experimental unit in a nested structure (families within populations). The plots were blocked by their position on the slope.

We selected three populations from each of the trial sites (Arenas, central Spain; Oriá, southeast Spain; Mimizan, southwest coast of

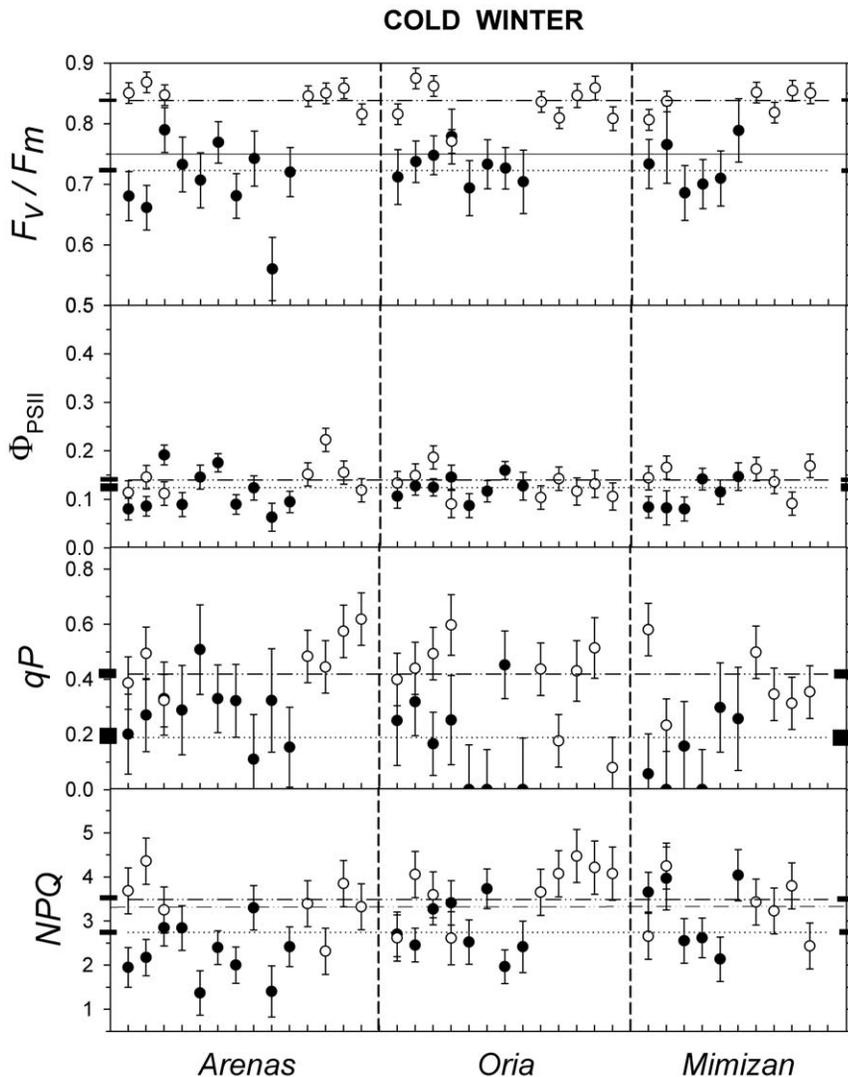


Figure 4. Family variation in the photochemical parameters during the cold winter. Least squared means \pm standard errors of the photochemical parameters for families from the *Arenas*, *Oria* and *Mimizan* populations present in the xeric (black circles) and mesic (white circles) trial sites in the cold winter. Families present at both sites are represented at the same position on the X axis. Abbreviations are the same as those in Fig. 3. doi:10.1371/journal.pone.0028772.g004

France) to test the photochemical parameters during the winters of 2006 and 2007. For the freeze test, we added the Moroccan *Tamrabta* population and measured 4 populations from the continental site (*XE*) exclusively. Location and climatic data for the trial sites and populations are presented in Figure 2 and Table 1.

Winter stress

At *XE*, physiological measurements were performed, contingent on the availability of plant material. Weather conditions limited the number of measurements at the *ME*. We collected data from 3 populations, evaluating 6 to 10 families per population and 4 to 8 trees per family. On each day of measurements, families and populations were selected at random in the field.

On clear and sunny winter days, photosynthetically active radiation (PAR) was $2198 \pm 23 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ at the *XE* and $2099 \pm 17 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ at the *ME*. Chlorophyll fluorescence was measured between 10:00 a.m. and 1:00 p.m. (13:00, solar time) for three consecutive days at the *XE*, followed

by another three consecutive days at the *ME*, with a modulated portable fluorescence monitoring system (Hansatech Instruments, FMS 2). Needles were illuminated with light from the FMS 2. PAR was adjusted to $1500 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ at the *XE* and $1360 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ at the *ME* to avoid photoinhibition. A portable computer was coupled to the FMS 2 to record any changes in the kinetics of the modulated chlorophyll fluorescence.

The photochemical parameters, Φ_{PSII} , F_v/F_m and q_p , and the photoprotective mechanism NPQ , provide valuable information about the electron transport system and the conversion and dissipation of the excess excitation energy into heat and can be used as indicators of plant stress [14]. Φ_{PSII} and F_v/F_m reflect the actual and maximum potential efficiency of the excitation energy captured by open PSII centers, respectively. While Φ_{PSII} indicates the proportion of absorbed energy that is used in photochemistry, the photochemical quenching, q_p , indicates the proportion of PSII reaction centers that are open and shows the level of fluorescence quenching due to the rapidly relaxing redox state of the primary quinone of PSII, Q_A . In contrast, the non-photochemical

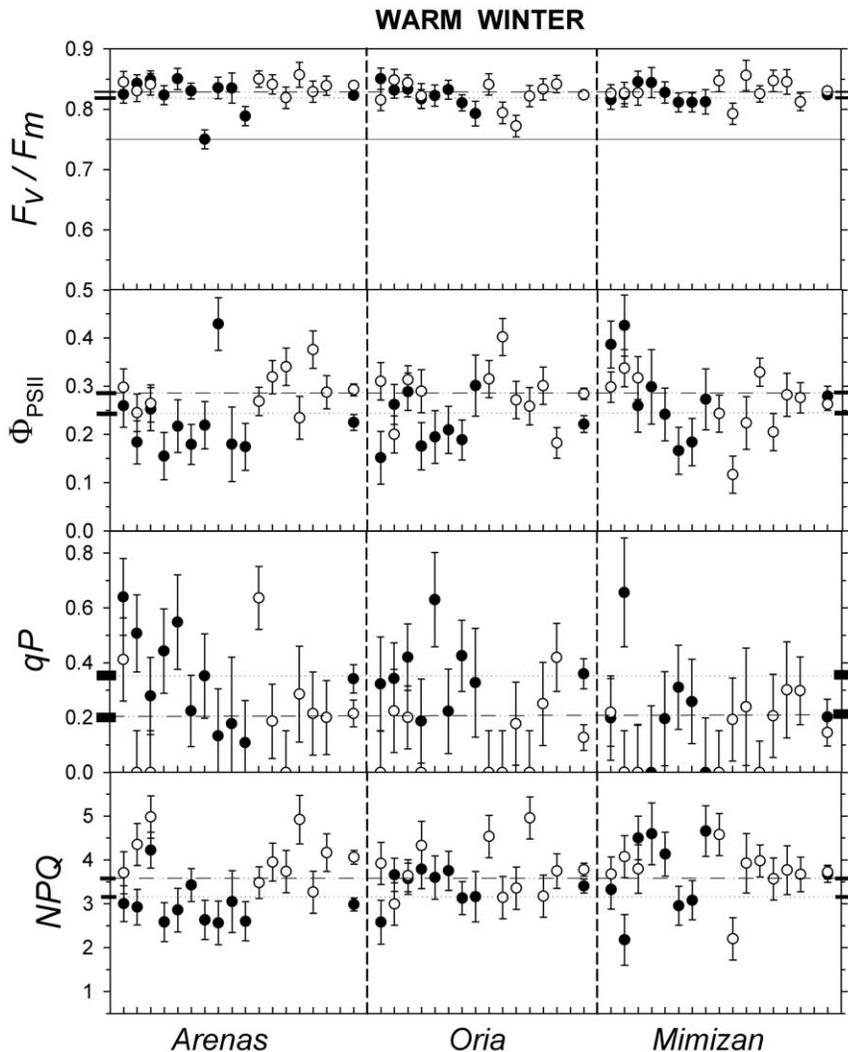


Figure 5. Family variation in photochemical parameters during the warm winter. Least squared means \pm standard errors of the photochemical parameters for the families from the *Arenas*, *Oria* and *Mimizan* populations present in the xeric (black circles) and mesic (white circles) trial sites in the warm winter. Families present at both sites are represented at the same position on the X axis. Abbreviations are the same as those in Fig. 3. The Y-axis scale is the same as that in Fig. 4 to highlight the differences between the winters. doi:10.1371/journal.pone.0028772.g005

quenching, NPQ , shows the level of fluorescence quenching due to the slowly relaxing high energy status of the thylakoids, i.e., the ΔpH -dependent processes that lower the efficiency of PSII as the rates of electron transport and carbon metabolism reach saturation at high photon fluxes. NPQ is directly proportional to the rate constant for the energy dissipation as heat.

The experimental protocol was essentially as that described by Genty et al. [40], with some modifications [41]. Several needles from the current year were dark-adapted using leaf clips provided with the FMS 2 and were kept in darkness for 30 min before estimating the minimum (F_0) and maximum (F_m) chlorophyll fluorescence at predawn. F_0 was measured by switching on the modulated light at 0.6 kHz. F_m and F'_m were measured at 20 kHz with a pulse of 6000 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ of white light for 1 s. F_0 and F'_0 were measured in the presence of far-red light (7 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) to completely oxidize the PSII acceptor side [41]. The actual fluorescence level (F_s) was obtained when fluorescence reached a steady-state value, which was verified visually on the computer screen. After the saturating pulse of light

for determining the maximum fluorescence in the light (F'_m), the fluorescence signal was allowed to re-equilibrate. When the signal reached the same steady-state level observed prior to the saturating pulse of light, it was monitored for approximately 20 s to attain the F'_0 level. The maximal photochemical efficiency of PSII, F_v/F_m , measured in the dark-adapted leaves, was given by the equation $(F_m - F_0)/F_m$ [42]. Actual PSII efficiency (Φ_{PSII}) was estimated as $(F'_m - F_s)/F'_m$ [43]. Photochemical quenching (qP) was estimated as $(F'_m - F_s)/F'_0$ [42]. Non-photochemical quenching (NPQ) was estimated as $(F_m - F'_m) - 1$ according to Bilger and Björkman [44].

Water potential

Predawn water potential, an indicator of plant water status at the moment of measurement, was obtained with a Scholander pressure chamber. Shoots from the outer parts of the crowns of ten individuals were taken from trees on the upper, middle and lower part of the slope. Shoots were wrapped in plastic film, cut with pruning shears and measured immediately.

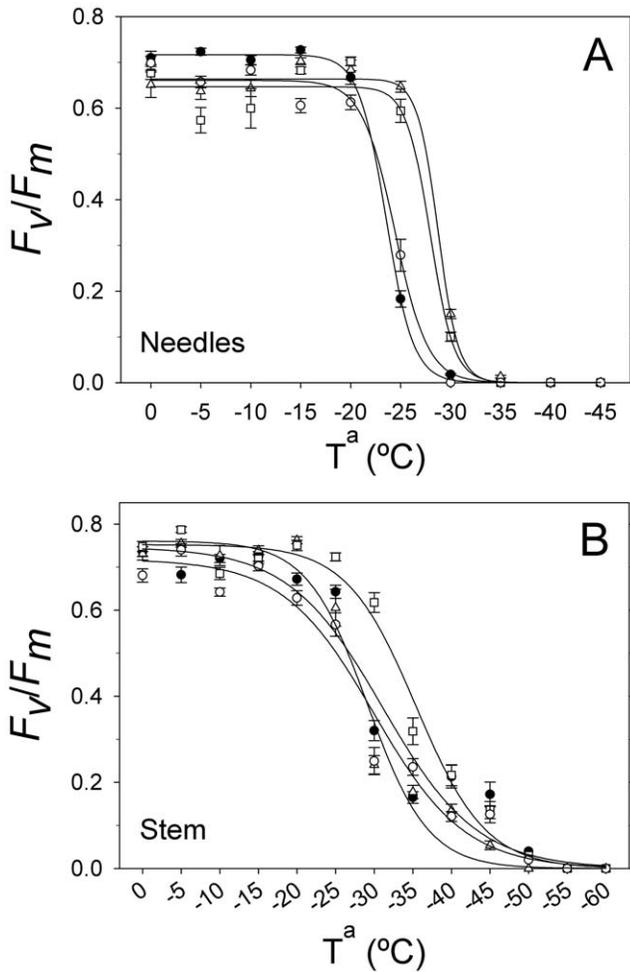


Figure 6. Population variation in the freezing tolerance of needles and stems. The relationship between the temperature ($^{\circ}\text{C}$) and maximum potential PSII efficiency (F_v/F_m) in the needles (A) and stems (B) of *Pinus pinaster* populations is shown: Oriá (filled circles), Mimizan (open circles), Tamrabta (squares) and Arenas (triangles). Error bars indicate standard errors of the mean values. doi:10.1371/journal.pone.0028772.g006

Freeze test

At the XE, branches from ten individuals per population were randomly selected in the field for each freezing temperature. We used current-year needles and branches from the three populations tested for winter stress and also added a new population, *Tamrabta*, to cover a wider range of winter temperatures and altitudes from the provenances. Immediately after collection, the samples were stored in a black bag inside a thermoelectric refrigerator at 4°C and brought to the laboratory for measurements. The study was conducted during February 2007.

We adapted a commercial freezer to obtain the temperature profiles. Cooling rates were established through an industrial controller (PMA Prozess-und Maschinen-Automation GmbH Mod. KS90, Germany) acting on a heating block with forced convection. Freezing was provided by the continuous performance of the freezer engine. Thermal homogeneity was achieved through a microfan system inside the chamber. This device provided a thermal stability of $\pm 0.1^{\circ}\text{C}$ onset temperature along the whole thermal profile. The samples (10 twigs per test temperature and population) were exposed to various freezing temperatures

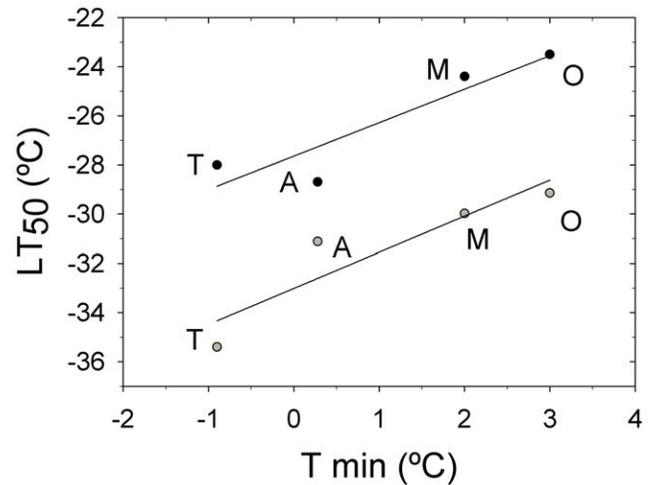


Figure 7. Populations ranked according to LT_{50} . The relationship between the LT_{50} of the needles (black circles) or stems (grey circles) and the mean minimum temperature of the coldest month (T_{min}) for the population origin is depicted. T: *Tamrabta*, A: *Arenas*, O: *Oriá*, M: *Mimizan*. doi:10.1371/journal.pone.0028772.g007

between 0 and -25°C . The rate of cooling was 5°C h^{-1} , and the samples were exposed to the freezing temperature for 1 h. This device was not able to achieve temperatures below -25°C , so temperature profiles between -30°C and -60°C were generated with a commercial freezer (CHF 250/80, Ingenieria de Climas, Barcelona, Spain) equipped with a PID digital temperature controller. The same experimental procedure described above was used for the exposition time and rate of cooling. The control temperature was 4°C . The freezing tolerance of the needles and stems (cortical bark chlorenchyma) was assessed by means of chlorophyll fluorescence. Curves were fitted to a logistic sigmoid function by regression analysis. LT_{50} was estimated as the inflection point of the sigmoid function:

$$y = a / (1 + e^{-(x - x_o)/b})$$

where x_o is the exposure temperature, y is the value of the variable used for the estimation of freezing tolerance, a defines the asymptote of the function, and b is the slope at the inflection point x .

Statistical analysis

To characterize the environments of the populations and testing sites, we constructed a dummy index based on climatic data,

Table 5. Correlations among photochemical parameters.

Parameter	Φ_{PSII}	qP	NPQ
F_v/F_m	0.24***	0.14***	0.49***
Φ_{PSII}	0	0.87***	-0.09*
qP	0	0	0.08*

Abbreviations are the same as those in Table 2. Bold numbers denote significant values. Significance levels: *** $P < 0.001$, ** $P < 0.01$, * $0.01 < P < 0.05$.

doi:10.1371/journal.pone.0028772.t005

Table 6. Correlations among the photochemical parameters, separated by winter and site.

Winter	Parameter	Xeric site			Mesic site		
		Φ_{PSII}	qP	NPQ	Φ_{PSII}	qP	NPQ
Cold	F_v/F_m	0.53***	0.46***	0.62***	0.46***	0.43***	0.43***
Cold	Φ_{PSII}	1	0.79***	0.08	1	0.93***	-0.08
Cold	qP	0.79***	1	-0.25**	0.93***	1	0.05
Warm	F_v/F_m	0.14	0.04	0.31***	0.08	-0.04	0.32***
Warm	Φ_{PSII}	1	0.85***	-0.27**	1	0.77***	-0.10
Warm	qP	0.85***	1	0.11	0.77***	1	-0.24**

Abbreviations are the same as those in Table 2. Bold numbers denote significant values. Signification level:

*** $P < 0.001$,

** $P < 0.01$,

* $0.01 < P < 0.05$.

doi:10.1371/journal.pone.0028772.t006

including as much variability as possible. Using a GIS (Geographic Information System), we extracted values for the 19 bioclimatic variables provided by the “Worldclim” model [45]. For each variable, we calculated the mean value for a circular surface with a 10 km radius centered on the coordinates of the testing sites and population provenances. The climatic index used for comparison purposes was the first component value obtained from a principal component analysis (PCA) of these bioclimatic data.

According to the experimental design, and on the basis of the subsampling that was performed, a set of mixed models was used for all variables. The normality and homoscedasticity of the data were confirmed.

The general model established was as follows:

$$y_{ijklm} = \mu + s_i + l_j + s_i \times l_j + P_k + P_k \times s_i + P_k \times l_j + P_k(s_i \times l_j) + f_l(P_k) + f_l(P_k \times s_i) + f_l(P_k \times l_j) + f_l(P_k \times s_i \times l_j) + \varepsilon_{ijklm}$$

where

y_{ijklm} is the value of the variable for the m_{th} seedling from the k_{th} population within the l_{th} family measured at the j_{th} site during the i_{th} winter;

μ is the overall mean of the variable;

s_i is the effect of the i_{th} winter ($i = 1-2$);

l_j is the effect of the j_{th} site ($j = 1-2$);

P_k is the effect of the k_{th} population ($k = 1-3$);

f_l is the effect of the l_{th} family ($l = 1-10$) within the k_{th} population;

ε_{ijklm} is the residual ($m = 1-4$).

This model was applied both generally and individually, by winter and site, removing the sources of variation, “site” and “winter”, from the model. The model was analyzed as a mixed model with fixed (site, winter, population and family) and random (error) effects, and the components of variance were obtained by restricted maximum likelihood (REML). The best linear unbiased estimators and predictors (BLUE and BLUP) for fixed and random factors, respectively, were obtained using SAS [46].

Pearson correlation coefficients were obtained to analyze the relationships between the variables considered.

Table 7. Percentages of variance (%).

Sources of variation	F_v/F_m	Φ_{PSII}	qP	NPQ
Site	0	0	0	5
Winter	4	30	38	14
Site x winter	49	2	4	3
Pop	0	0	0	0
Pop x site	0	0	0	0
Pop x winter	0	0	0	0
Pop x (site x winter)	0	0	0	0
Fam (Pop)	0	0	0	0
Fam (Pop) x site	1	0	5	0
Fam (Pop) x winter	0	2	3	0
Fam (Pop) x (site x winter)	1	8	9	11
Error	45	57	41	66

Abbreviations are the same as those in Table 2. Empty boxes denote zero values.

doi:10.1371/journal.pone.0028772.t007

Table 8. Percentages of variance by site and winter (%).

Parameter variation	Sources of variation	Cold winter		Warm winter	
		Site XE	Site ME	Site XE	Site ME
F_v/F_m	Pop	0	0	0	1
F_v/F_m	Fam (Pop)	1	3	8	2
F_v/F_m	residual	84	12	44	45
Φ_{PSII}	Pop	0	0	0	0
Φ_{PSII}	Fam (Pop)	12	7	12	9
Φ_{PSII}	residual	42	40	52	26
qP	Pop	0	0	3	0
qP	Fam (Pop)	11	2	17	25
qP	residual	57	30	30	25
NPQ	Pop	4	0	3	0
NPQ	Fam (Pop)	7	5	6	7
NPQ	residual	41	43	45	40

Abbreviations are the same as those in Table 2.

doi:10.1371/journal.pone.0028772.t008

Author Contributions

Conceived and designed the experiments: LC EG-P EN. Performed the experiments: LC. Analyzed the data: EN. Contributed reagents/materials/analysis tools: LC EG-P EN. Wrote the paper: LC.

References

- Zas R, Merlo E, Fernandez-Lopez J (2004) Juvenile - Mature genetic correlations in *Pinus pinaster* Ait. under different nutrient x water regimes. *Silvae Genetica* 53: 124–129.
- Alia R, Moro J, Denis JB (1997) Performance of *Pinus pinaster* provenances in Spain: interpretation of the genotype by environment interaction. *Canadian Journal of Forest Research* 27: 1548–1559.
- Corcuera L, Gil-Pelegrin E, Notivol E (2010) Phenotypic plasticity in *Pinus pinaster* $\delta^{13}\text{C}$: environment modulates genetic variation. *Annals of Forest Science* 67: 812.
- Corcuera L, Cochard H, Gil-Pelegrin E, Notivol E (2011) Phenotypic plasticity in mesic populations of *Pinus pinaster* improves resistance to xylem embolism (P_{50}) under severe drought. *Trees* 25: 1033–1042.
- Corcuera L, Gil-Pelegrin E, Notivol E (2012) Aridity promotes differences in proline and phytohormone levels in *Pinus pinaster* populations from contrasting environments. *Trees* (in press).
- Major JE, Johnsen KH (1996) Family variation in photosynthesis of 22-year-old black spruce: A test of two models of physiological response to water stress. *Journal of Forest Research* 26: 1922–1933.
- Lamontagne M, Margolis H, Bigras F (1998) Photosynthesis of black spruce, jack pine, and trembling aspen after artificially induced frost during the growing season. *Canadian Journal of Forest Research* 28: 1–12.
- Lamontagne M, Bigras FJ, Margolis HA (2000) Chlorophyll fluorescence and CO_2 assimilation of black spruce seedlings following frost in different temperature and light conditions. *Tree Physiology* 20: 249–255.
- Adams WW, Demmig-Adams B (1994) Carotenoid composition and down regulation of photosystem II in three conifer species during the winter. *Physiologia Plantarum* 92: 451–458.
- Egerton JJ, Banks JC, Gibson A, Cunningham RB, Ball MC (2000) Facilitation of seedling establishment: reduction in irradiance enhances winter growth of *Eucalyptus pauciflora*. *Ecology* 81: 1437–1449.
- Larcher W (2000) Temperature stress and survival ability of Mediterranean sclerophyllous plants. *Plant biosystems* 134: 279–295.
- Corcuera L, Morales F, Abadía A, Gil-Pelegrin E (2005) Seasonal changes in photosynthesis and photoprotection in a *Quercus ilex* subsp. *ballota* woodland located in its upper altitudinal extreme in the Iberian Peninsula. *Tree Physiology* 25: 599–608.
- Corcuera L, Morales F, Abadía A, Gil-Pelegrin E (2005) The effect of low temperatures on the photosynthetic apparatus of *Quercus ilex* subsp. *ballota* at its lower and upper altitudinal limits in the Iberian Peninsula and during a single freezing-thawing cycle. *Trees Structure and Function* 19: 99–108.
- Maxwell K, Johnson GN (2000) Chlorophyll fluorescence - a practical guide. *Journal of Experimental Botany* 51: 659–668.
- Adams III WW, Demmig-Adams B (1995) The xanthophyll cycle and sustained thermal energy dissipation in *Vinca minor* and *Euonymus kiautschovicus* in winter. *Plant Cell and Environment* 18: 117–127.
- Ladjal M, Epron D, Ducrey M (2000) Effects of drought preconditioning on the thermotolerance of photosystem II and susceptibility of photosynthesis to heat stress in cedar seedlings. *Tree Physiology* 20: 1235–1241.
- Verhoeven AS, Demmig-Adams B, Adams III WW (1997) Enhanced employment of the xanthophyll cycle and thermal energy dissipation in spinach exposed to high light and N stress. *Plant Physiology* 113: 817–824.
- Epron D, Dreyer E, Breda N (1992) Photosynthesis of oak trees (*Quercus petraea*) during drought under field conditions: Diurnal course of net CO_2 assimilation and photochemical efficiency of photosystem II. *Plant Cell and Environment* 15: 809–820.
- Benowicz A, Hirondele SL, El-Kassaby YA (2001) Patterns of genetic variation in mountain hemlock (*Tsuga mertensiana* (Bong.) Carr.) with respect to height growth and frost hardiness. *Forest Ecology and Management* 154: 23–33.
- Lu PX, Joyce DG, Sinclair RW (2003) Geographic variation in cold hardiness among eastern white pine (*Pinus strobus* L.) provenances in Ontario. *Forest Ecology and Management* 178: 329–340.
- Lindgren K, Hällgren JE (1993) Cold-acclimation of *Pinus contorta* and *Pinus sylvestris* assessed by chlorophyll fluorescence. *Tree Physiology* 13: 97–106.
- Koehn AC, Roberds JH, Doudrick RL (2003) Variation among slash pine families in chlorophyll fluorescence traits. *Canadian Journal of Forest Research* 33: 1102–1109.
- Peguero-Pina JJ, Morales F, Flexas J, Gil-Pelegrin E, Moya I (2008) Photochemistry, remotely-sensed physiological reflectance index (PRI) and deoxidation state of the xanthophyll cycle in *Quercus coccifera* under intense drought. *Oecologia* 156: 1–11.
- Fracheboud Y, Ribaut JM, Vargas M, Messmer R, Stamp P (1999) Chlorophyll fluorescence as a selection tool for cold tolerance of photosynthesis in maize (*Zea mays* L.). *Journal of Experimental Botany* 50: 1533–1540.
- Ögren E (1999) Fall frost resistance in willows used for biomass production. I. Characterization of seasonal and genetic variation. *Tree Physiology* 19: 749–754.
- Major JE, Barsi DC, Mosseler A, Campbell M, Rajora OP (2003) Light-energy processing and freezing-tolerance traits in red spruce and black spruce: species and seed-source variation. *Tree Physiology* 23: 685–694.
- Medlyn BE, Loustau D, Delzon S (2002) Temperature response of parameters of a biochemically based model of photosynthesis. I. Seasonal changes in mature maritime pine (*Pinus pinaster* Ait.). *Plant Cell and Environment* 25: 1155–1165.
- Baquedano FJ, Castillo FJ (2006) Comparative ecophysiological effects of drought on seedlings of the Mediterranean water-saver *Pinus halepensis* and water-spenders *Quercus coccifera* and *Quercus ilex*. *Trees Structure and Function* 20: 689–700.
- Baquedano FJ, Castillo FJ (2007) Drought tolerance in the Mediterranean species *Quercus coccifera*, *Quercus ilex*, *Pinus halepensis*, and *Juniperus phoenicea*. *Photosynthetica* 45: 229–238.
- Baquedano FJ, Valladares F, Castillo FJ (2008) Phenotypic plasticity blurs ecotypic divergence in the response of *Quercus coccifera* and *Pinus halepensis* to water stress. *European Journal of Forest Research* 127: 495–506.
- Aranda I, Castro L, Alia R, Pardos JA, Gil L (2005) Low temperature during winter elicits differential responses among populations of the Mediterranean evergreen cork oak (*Quercus suber*). *Tree Physiology* 25: 1085–1090.
- Prieto P, Peñuelas J, Llusia J, Asensio D, Estiarte M (2009) Effects of long-term experimental night-time warming and drought on photosynthesis, Fv/Fm and stomatal conductance in the dominant species of a Mediterranean shrubland. *Acta Physiologia Plantarum* 31: 729–739.
- Weng JH, Lai KM, Liao TS, Hwang MY, Chen YN (2009) Relationships of photosynthetic capacity to PSII efficiency and to photochemical reflectance index of *Pinus taiwanensis* through different seasons at high and low elevations of sub-tropical Taiwan. *Trees Structure and Function* 23: 347–356.
- Soukupova J, Csefalvay L, Urban O, Kosvancova M, Marek M, et al. (2008) Annual variation of the steady-state chlorophyll fluorescence emission of evergreen plants in temperate zone. *Functional Plant Biology* 35: 63–76.
- Warren CR, Tausz M, Adams MA (2005) Does rainfall explain variation in leaf morphology and physiology among populations of red ironbark (*Eucalyptus sideroxylon* subsp. *tricarpa*) grown in a common garden?. *Tree Physiology* 25: 1369–1378.
- Colom MR, Prato EP, Giannini R (2003) Chlorophyll fluorescence and photosynthetic response to light in 1-year-old needles during spring and early summer in *Pinus leucodermis*. *Trees Structure and Function* 17: 207–210.
- Lopez R, Rodriguez-Calcerrada J, Gil L (2009) Physiological and morphological response to water deficit in seedlings of five provenances of *Pinus canariensis*: potential to detect variation in drought-tolerance. *Trees Structure and Function* 23: 509–519.
- Climent J, Costa e Silva F, Chambel R, Pardos M, Almeida H (2009) Freezing injury in primary and secondary needles of Mediterranean pine species of contrasting ecological niches. *Annals of Forest Science* 66: 407.
- Alia R, Gil L, Pardos JA (1995) Performance of 43 *Pinus pinaster* Ait. Provenances on 5 locations in central Spain. *Silvae Genetica* 44: 75–81.
- Genty B, Briantais JM, Baker NR (1989) The relationship between quantum yield of photosynthetic electron transport and quenching of chlorophyll fluorescence. *Biochimica et Biophysica Acta* 990: 87–92.
- Belkhodja R, Morales F, Quílez R, López-Millán AF, Abadía A, Abadía J (1998) Iron deficiency causes changes in chlorophyll fluorescence due to the reduction in the dark of the photosystem II acceptor side. *Photosynthesis Research* 56: 265–276.
- Van Kooten O, Snel JFH (1990) The use of chlorophyll fluorescence in plant stress physiology. *Photosynthesis Research* 25: 147–150.
- Harbison J, Genty B, Baker NR (1989) Relationship between the quantum efficiencies of photosystems I and II in pea leaves. *Plant Physiology* 90: 1029–1034.
- Bilger W, Björkman O (1990) Role of the xanthophyll cycle in photoprotection elucidated by measurements of light-induced absorbance changes, fluorescence and photosynthesis in leaves of *Hedera canariensis*. *Photosynthesis Research* 25: 173–185.
- Hijmans RJ, Cameron SE, Parra JL, Jones PG, Jarvis A (2005) Worldclim: High resolution interpolated surfaces for global land areas. *International Journal of Climatology* 25: 1965–1978.
- SAS (1999) SAS OnlineDoc version eight. SAS, Institute Inc., Cary, NC, USA.